

A New Deployment of the Burrows-Wheeler Transform to Improve DNA Methylation Analysis



Ricardo Olanda Rodríguez
Mariano Pérez Martínez
Juan M. Orduña Huertas
César González Segura

Dpto. de Informática, Universidad de Valencia
Juan.orduna@uv.es

Introduction

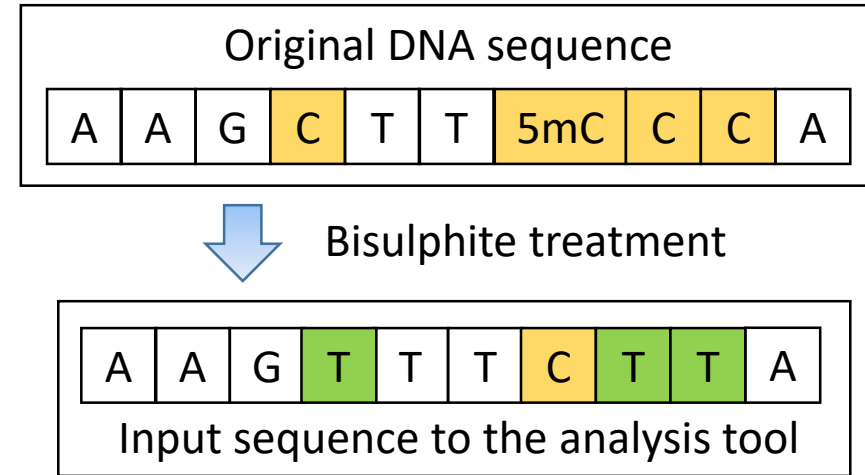
- The introduction of Next Generation Sequencing (NGS) has increased the size and length of DNA samples.
- Current sequencers produce billions of short DNA samples, with lengths usually surpassing 400 nt^[1].
- Upcoming sequencers will be able to generate samples with lengths up to thousands of nts, producing huge datasets as a result ^[2].



* [Illumina NextSeq 500 (https://research.ncsu.edu/gsl/files/2015/08/Illumina_NextSeq_500.jpg)]

DNA Methylation

- Methylation is a particular topic of DNA analysis, relevant in the study of cancer, diabetes and other diseases.
- Through bisulfite-treatment, the methylation status can be retrieved with base-pair resolution.
- Several methylation analysis tools have been developed^[3,4,5,6], however their performance decreases with read lengths over 150 nts.

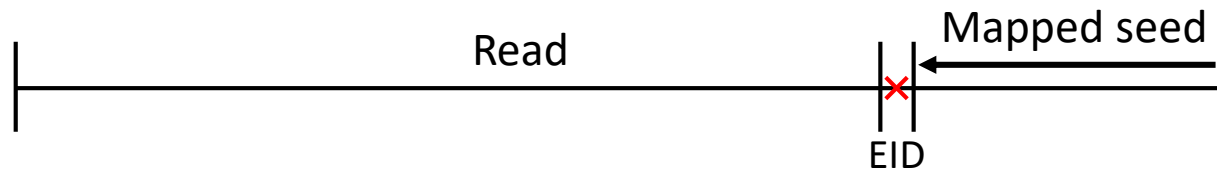


Methylation Analysis Pipeline

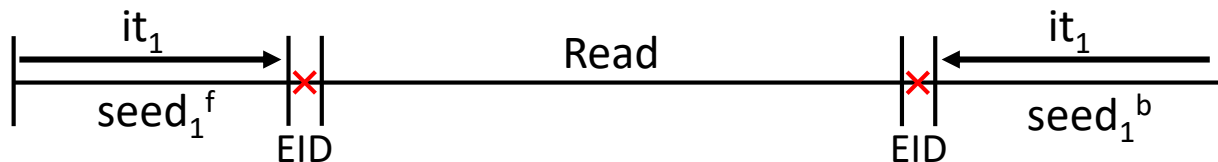
- Our approach is based on using two mapping algorithms, the Burrows Wheeler Transform^[7] and the Smith & Waterman Algorithm^[8].
- In HPG-Methyl 2, we propose a new strategy to reduce the execution time and increase sensitivity even for long reads using two techniques:
 - A bidirectional BWT, which can discard wrong alignments more efficiently.
 - A new parallel pipeline scheme, reducing the number of invalid candidate regions and the execution time of the SWA.

New Implementation of the Burrows-Wheeler Transform

- The Burrows-Wheeler transform can be used as a backward search method, allowing errors, insertions or deletions (EIDs)^[7].
- Unidirectional BWT performs a backward search, mapping a read segment to the reference genome base by base, until an EID is found.



- Bidirectional BWT performs the search from both ends simultaneously, doubling the maximum number of allowed EIDs.

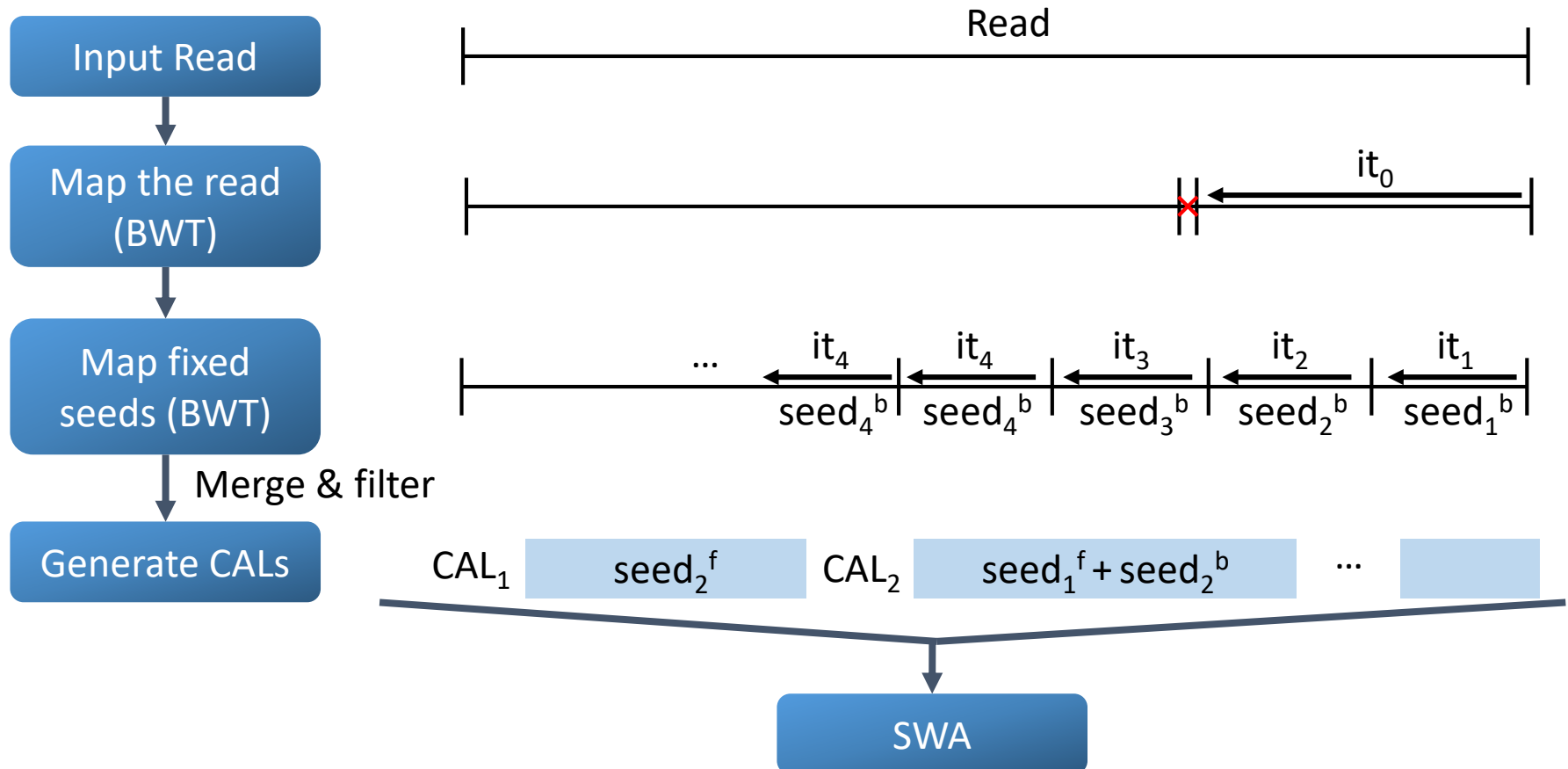


Implementation in a New Parallel Pipeline

- We have used HPG-Methyl^[9], a parallel methylation analysis tool, as the basis to implement these new strategies.
- To take full advantage of the new BWT implementation, HPG-Methyl features several changes to the pipeline:
 - Several stages are merged into a single, more flexible stage.
 - This new stage produces fewer candidate regions (CALs), but much more effective.
 - As a result, use of the SWA algorithm is greatly reduced, improving execution times.

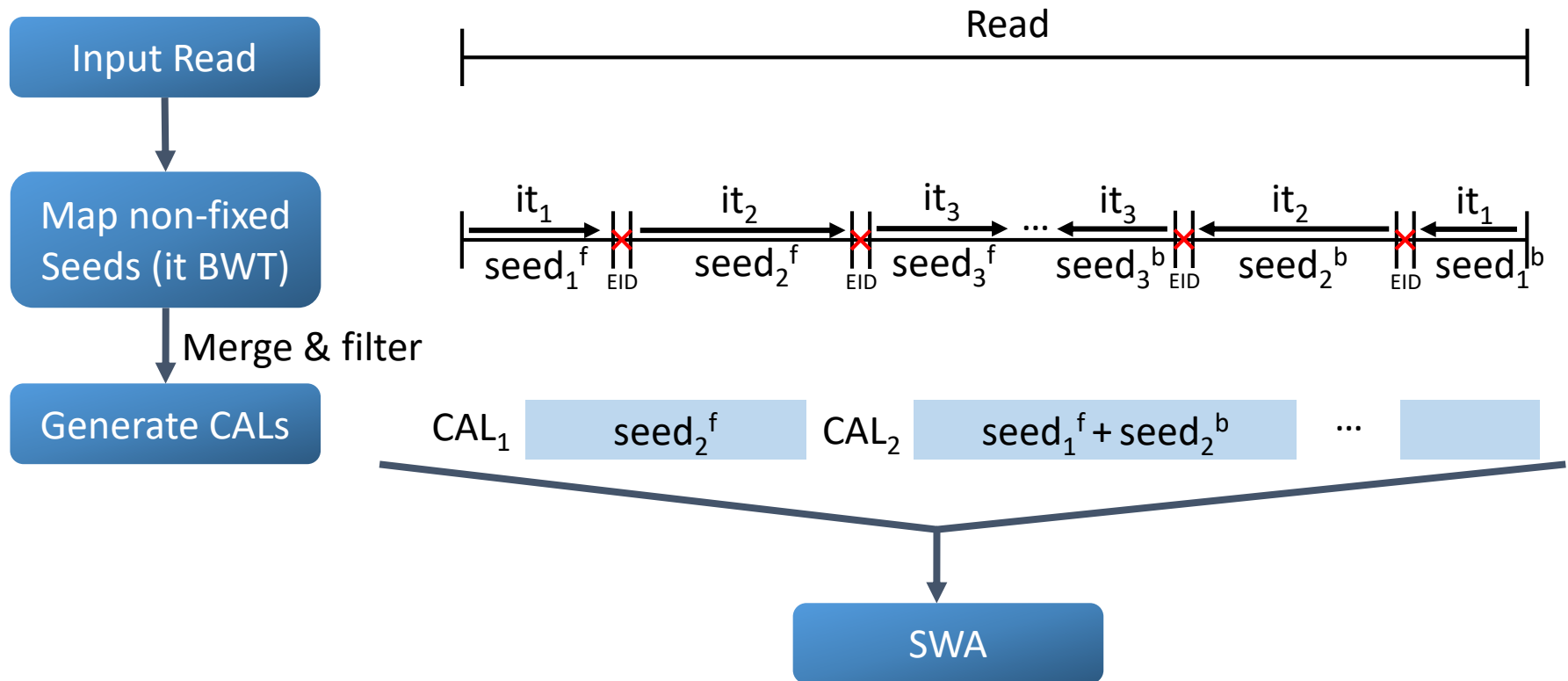
Implementation in a New Parallel Pipeline

- Stages of the original pipeline in HPG-Methyl.



Implementation in a New Parallel Pipeline

- Modified stages of the parallel pipeline in HPG-Methyl 2.



Performance Evaluation

The performance of HPG-Methyl 2 was compared with HPG-Methyl^[9] and Bismark^[3], using:

- Synthetic methylation datasets with read lengths between 75 nt and 3200 nt, with 4 million reads.
- Real methylation datasets from the European Nucleotide Archive^[10].

Tests were conducted in a computer platform as in the table, measuring:

- Sensitivity.
- Execution time.

Computer Platform
Intel i7 3930K (6 cores @ 3,8 GHz)
48 GB RAM
SSD Storage
Ubuntu 14.04 LTS

Performance Evaluation: sensitivity

- HPG-Methyl 2 yields the most accurate results, with sensitivity rates over 99% in all cases.

Length (nt)	HPG-Methyl 2		HPG-Methyl		Bismark	
	Right	Wrong	Right	Wrong	Right	Wrong
75	99,50%	0,69%	93,37%	0,62%	88,30%	0,10%
150	99,01%	0,46%	96,87%	0,80%	94,59%	0,08%
400	99,75%	0,18%	97,55%	0,48%	97,55%	0,10%
800	99,93%	0,06%	96,94%	0,48%	98,45%	0,08%
1600	99,75%	0,06%	96,94%	0,48%	—*	—*
3200	99,68%	0,08%	96,42%	0,49%	—*	—*

* Aborted after waiting for 3 days (4320 minutes)

Performance Evaluation: execution time

- Our new implementation allows for a significant reduction in the execution time.
- Now HPG-Methyl scales with the increasing sequence lengths of NGS.

Length (nt)	HPG-Methyl 2	HPG-Methyl	Bismark
75	1,288	1,366	69,579
150	1,550	1,950	106,173
400	5,041	10,85	248,107
800	11,26	50,60	1246,89
1600	34,44	996,567	—*
3200	164,593	7733,38	—*

* Aborted after waiting for 3 days (4320 minutes)

Conclusions

- We have proposed a new strategy for methylation analysis, using a bidirectional BWT implementation and a new parallel pipeline.
- Our implementation reduces execution times by an order of magnitude and achieves sensitivity rates over 99%, even for very long reads.
- The software is open source and available in Github:
<https://github.com/grev-uv/hpg-methyl>

Future Work

- This strategy can be exported to other methylation analysis tools, and to DNA and RNA analysis tools.
- We plan on applying the same strategy of bidirectional BWT to similar tools, such as HPG-Aligner.
- Also, HPG-Methyl will keep being updated with performance improvements and new features.

References

- [1] Inc. PACBIO Pacific Biosciences of California, “PACBIO RS II: The original long-read sequencer,” <http://www.pacb.com/products-and-services/pacbio-systems/rsii/>, 2016.
- [2] Joaquin Tarraga, Asunción Gallego, Vicente Arnau, Ignacio Medina, and Joaquin Dopazo, “Hpg pore: an efficient and scalable framework for nanopore sequencing data.,” *BMC bioinformatics*, vol. 17, pp. 107, 2016 2016.
- [3] Felix Krueger and Simon R. Andrews, “Bismark: a flexible aligner and methylation caller for bisulfite-seq applications,” *Bioinformatics*, vol. 27, no. 11, pp. 1571–1572, 2011.
- [4] Pao-Yang Chen, Shawn Cokus, and Matteo Pellegrini, “Bs seeker: precise mapping for bisulfite sequencing.,” *BMC Bioinformatics*, vol. 11, pp. 203, 2010.
- [5] Yuanxin Xi and Wei Li, “BSMAP: whole genome bisulfite sequence MAPping program.,” *BMC bioinformatics*, vol.10, no. 1, pp. 232+, 2009.
- [6] Yuanxin Xi, Christoph Bock, Fabian Maller, Deqiang Sun, Alexander Meissner, and Wei Li, “Rrbsmap: a fast, accurate and user-friendly alignment tool for reduced representation bisulfite sequencing.,” *Bioinformatics*, vol. 28, no. 3, pp. 430–432, 2012.
- [7] H. Li and R. Durbin, “Fast and accurate short read alignment with burrows-wheeler transform.,” *Bioinformatics*, vol. 25, no. 14, pp. 1754–60, 2009.
- [8] T. F. Smith and M. S. Waterman, “Identification of common molecular subsequences.,” *Journal of molecular biology*, vol. 147, no. 1, pp. 195–197, Mar. 1981.
- [9] Joaquín Tárraga, Mariano Pérez, Juan M Orduña, José Duato, Ignacio Medina, and Joaquín Dopazo, “A parallel and sensitive software tool for methylation analysis on multicore platforms.,” *Bioinformatics (Oxford, England)*, vol. 31, no. 19, pp. 3130–3138, 2015 Jun 10 2015.
- [10] <http://www.ebi.ac.uk/ena/data/view/SRR309230> and <http://www.ebi.ac.uk/ena/data/view/SRR837425>

A New Deployment of the Burrows-Wheeler Transform to Improve DNA Methylation Analysis



Ricardo Olanda Rodríguez
Mariano Pérez Martínez
Juan M. Orduña Huertas
César González Segura

Dpto. de Informática, Universidad de Valencia
Juan.orduna@uv.es