European Journal of Medicinal Chemistry 87 (2014) 834-842

Contents lists available at ScienceDirect

European Journal of Medicinal Chemistry

journal homepage: http://www.elsevier.com/locate/ejmech

Mini-review

Synthetic derivatives of aromatic abietane diterpenoids and their biological activities

Miguel A. González

Departamento de Química Orgánica, Universidad de Valencia, E-46100 Burjassot, Valencia, Spain

A R T I C L E I N F O

Article history: Received 26 August 2014 Received in revised form 6 October 2014 Accepted 9 October 2014 Available online 13 October 2014

Keywords: Abietane Diterpenoid Dehydroabietic acid Ferruginol Dehydroabietylamine Carnosic acid

1. Introduction

Natural products have played a dominant role in the drug discovery efforts for the treatment of human diseases [1]. Abietanes are a family of naturally occurring diterpenoids that have been isolated from a variety of terrestrial plant sources. During the last three decades, many new members of this family of natural products have been isolated and described in several specific reviews on naturally occurring diterpenoids by Professor Hanson [2]. These compounds exhibit a wide variety of interesting biological activities, which has generated significant interest from the medicinal and pharmacological communities. The biological activities of natural abietane acids and their derivatives have been reviewed up to 1992 [3]. In this review, our attention is focused on diterpenoids characterised by tricyclic structures having the abietane (**I**, C₂₀) (Fig. 1) carbon framework and an aromatic ring C.

Aromatic abietanes comprised the largest group of components of naturally occurring abietanes. They possess an aromatic ring C and a different degree of oxygenation at several positions. This group of abietanes is exemplified by dehydroabietic acid (**2**) and ferruginol (**3**) which were discovered more than seventy years ago [4,5]. Both structures were assigned based on chemical data. Dehydroabietic acid (**2**) was initially obtained from chemical

http://dx.doi.org/10.1016/j.ejmech.2014.10.023 0223-5234/© 2014 Elsevier Masson SAS. All rights reserved.

A B S T R A C T

Naturally occurring aromatic abietane diterpenoids (dehydroabietanes) exhibit a wide range of biological activities. A number of synthetic studies aimed at modifying the abietane skeleton in order to obtain new potential chemotherapeutic agents have been reported. In this study, the biological activities of synthetic derivatives of aromatic abietane diterpenoids are reviewed.

© 2014 Elsevier Masson SAS. All rights reserved.

studies starting from abietic acid (1), later, it was found in resin or extracts of conifers [6]. Ferruginol (3) was firstly isolated in 1939 from the resin of the Miro tree (*Podocarpus ferrugineus*), endemic to New Zealand [5]. Another typical aromatic abietane is carnosic acid (4), which is found in the popular Lamiaceae herbs, sage (*Salvia officinalis*) and rosemary (*Rosmarinus officinalis*) [7]. Dehydroabietylamine (5), an abietane diterpenic amine derived from abietic acid (1), is the main component of disproportionated rosin amine. The diterpenoid skeleton present in these abietanes has been the object of derivatisation in screening for new potential chemotherapeutic agents. In this study, the biological activities of dehydroabietic acid, ferruginol, carnosic acid and dehydroabietylamine derivatives are reviewed.

2. Biological activities of aromatic abietane derivatives

2.1. Dehydroabietic acid derivatives

Over the past few years, a lot of research has been committed to the synthesis of dehydroabietic acid (**2**, DHA) derivatives. Dehydroabietic acid (**2**) displays not only antiulcer and antimicrobial properties but also antitumour effects. Modifications in rings B and C as well as manipulation of the carboxyl group at C-18 of DHA (**2**) have been studied in order to enhance its properties. In one of the early studies aimed at searching for new antiulcer agents with a broad cytoprotective effect, Wada et al. prepared more than







E-mail address: Miguel.A.Gonzalez@uv.es.



Fig. 1. Abietane numbering system and abietanes 1-5.

seventy derivatives of DHA (2), introducing a hydrophilic residue (amino, carbamoyl, carbamate, ureide, sulfonyl, or sulfamoyl) onto the lipophilic dehydroabietane skeleton [8]. The antisecretory and antipepsin activities were evaluated as a preliminary evaluation of antiulcer activity. The results obtained showed that DHA (2) has a moderate antisecretory action (22% inhibition of secretion at an intraperitoneally dose of 30 mg/kg in rats) and had no antipepsin activity. Among the tested compounds, the salts of 12sulfodehydroabietic acid (6a-b) (Fig. 2) were found to exhibit remarkably high antipepsin activity (92–96% inhibition at 100 mg/ kg) without aldosterone-like activity shown by other antiulcer agents. Further research on the molecule 6a (12sulfodehydroabietic acid monosodium salt) has led to the development of the drug ecabet[®] (ecabet sodium) for the treatment of reflux oesophagitis and peptic ulcer disease. The gastroprotective and cytotoxic effect of a series of DHA (2) derivatives at C-18 has been reported [9]. In this study, DHA (2) presented a dose-related gastroprotective effect on HCl/EtOH-induced gastric lesions in mice (59% inhibition at 100 mg/kg). The aromatic amide derivatives **7a**–g (Fig. 2) showed a strong gastroprotective activity (67–85% inhibition of gastric lesions) with low cytotoxicity ($IC_{50} > 1000 \mu M$) for both fibroblasts (MRC-5) and human epithelial gastric cell line (AGS).

The antimicrobial activity of resin acid derivatives has been reviewed up to 2006, including some DHA (**2**) derivatives [10]. For example, C-13 deisopropylated compounds **8** and **9** (Fig. 3) were the most active inhibiting the growth of several filamentous fungi (*Actinomucor harzii, Cladosporium cucumerinum, Mucor racemosus, Rhizopus arrhizus, Rhizopus stolonifer,* and *Syncephalastrum racemosum*) and also the Gram-positive bacterium *Staphylococcus aureus* [11]. Both compounds did not inhibit the growth of Gramnegative bacteria, *Escherichia coli* and *Klebsiella pneumonia.* However, in combination, those two compounds (**8** and **9**) inhibited the growth of those organisms suggesting a synergistic effect. The presence of an aldehyde group, compounds **10a** and **10b** (Fig. 3),



Fig. 2. Antiulcer DHA derivatives 6-7.



Fig. 3. Antifungal DHA derivatives 8-12.

seemed to be important for the antiyeast activity against Candida albicans, Candida kruzei and Candida parapsilosis [12]. More recently, González et al. demonstrated that DHA (2) (MIC = $39.7 \,\mu g/$ mL against Aspergillus terreus) and dehydroabietane 11a (Fig. 3) (MIC = 50 and 63 μ g/mL against Aspergillus fumigates and Aspergillus niger, respectively) possess anti-Aspergillus activity [13]. These authors also described anti-Aspergillus activity for the phenol 11b (Fig. 3) (MIC = 25, 25 and 50 μ g/mL against A. fumigates, A. terreus and A. niger, respectively) [14]. A series of DHA (2) derivatives bearing 1.2.4-triazolo-thiazolidinone moieties, compounds **12a**-g (Fig. 3), have been synthesised and tested at 50 μ g/mL for antifungal activity against Fusarium oxysporum, Alternaria solani, Physalospora piricola, Cercospora arachidicola and Fusarium graminearum [15]. With the exception of 12c, all compounds were most effective against F. graminearum, being compound 12d the most potent with inhibition ratio of 70.9%. Compound 12c showed the greatest inhibition ratio of 51.9% against F. oxysporum, and compound 12e displayed the greatest inhibition ratio of 56.8% against *P. piricola*.

Other antimicrobial studies include the synthesis of some thioureas, compounds **13a–k** (Fig. 4), and the corresponding 1,2,4-triazolo-aniline derivatives **14a–f** (Fig. 4), which were tested against *Bacillus subtilis* and *E. coli* [16]. Compounds **13j**, **13e**, **13f** and **14b** possess antibacterial activity against *B. subtilis* at a test concentration (for **13j**: 50 mg/mL; for **13e**, **13f**, **14b**: 100 mg/mL) while compounds **13b**, **13h**, **13i** and **14e** possess antibacterial activity against *E. coli* at a test concentration of 100 mg/mL.

A series of novel dibenzo-carbazole derivatives of DHA (2), compounds 15a-m (Fig. 4), were synthesised and tested against four bacteria (B. subtilis, S. aureus, E. coli, and Pseudomonas fluorescens) and three fungi (Trichophyton rubrum, C. albicans and A. niger) [17]. Among the compounds tested, **15d**, **15e**, **15f** and **15m** exhibited pronounced antibacterial activities and 15e and 15m also showed moderate antifungal activities. Particularly, 15d exhibited stronger antibacterial activity against *B. subtilis* (MIC = $1.9 \mu g/mL$) comparable to positive control (amikacin, MIC = $0.9 \mu g/mL$). Later on, the same authors reported the synthesis and antimicrobial evaluation of N-substituted dibenzo-carbazole derivatives of DHA (2), compounds 16a-s (Fig. 4) [18]. Some of the synthesised compounds displayed pronounced antimicrobial activity against four bacteria (B. subtilis, S. aureus, E. coli, and P. fluorescens) with low MIC values ranging from 0.9 to 15.6 µg/mL. Among them, compounds 16j and 16r exhibited potent inhibitory activity comparable to reference drug amikacin. These authors have also described the synthesis and antibacterial evaluation of new N-acylhydrazone derivatives, compounds 17a-q (Fig. 4), from DHA (2) [19]. The compounds were evaluated against four microbial strains (B.



Fig. 4. Antimicrobial DHA derivatives 13-17.

subtilis, S. aureus, E. coli, and P. fluorescens). Among the tested compounds, compound **17p** (MIC = 1.9 μ g/mL) exhibited good antibacterial activity against S. aureus and B. subtilis comparable to positive control (amikacin, MIC = 0.9 μ g/mL). Compounds **17l**, **17n** and **17p** also showed strong inhibitions (3.9–7.8 μ g/mL) against the Gram-negative strains E. coli and P. fluorescens.

The antiviral activity of DHA (2) derivatives has also been studied. For example, Fonseca et al. described the synthesis of novel heterocycles (Fig. 5), such as benzimidazoles (**18a**–**g**), quinoxalines (**19a**–**c**), and indoles **20** and **21** as potential antiviral agents [20]. Biological evaluation showed that compounds 18b, 18e, 19c, and 21 inhibited both varicella-zoster virus (VZV) and cytomegalovirus (CMV) replication at a concentration ca. 5- to 10-fold lower than the cytotoxic concentration, when tested in human embryonic lung cells. The potencies of 18a, 18b, 18d, 18e, 19c, 20 and 21 as anti-VZV agents were comparable to that of acyclovir ($IC_{50} = 0.3 - 3.0 \,\mu g/mL$), while the potencies of 18a, 18b, 18e, 19c and 21 as anti-CMV agents were comparable to that of ganciclovir (IC₅₀ = $0.9-1.5 \ \mu g/mL$). However, all these derivatives were not active against herpes simplex virus (HSV-1 and HSV-2), vaccinia virus, vesicular stomatitis virus, Coxsackie viruss B4, respiratory syncytial virus, and human immunodeficiency virus at a concentration of 400 pg/mL. Two different studies confirmed anti-herpetic activity for DHA (2) derivatives. Firstly, Tagat et al. reported that A-ring hydroxylated compounds 22a and 22b (Fig. 5) show HSV-2 inhibitory activity and



Fig. 5. Antiviral DHA derivatives 18-23.

concluded that they interfere with an early event in viral replication [21]. And secondly, Gonzalez et al. described that dehydroabietinol acetate **23** (Fig. 5) showed significant anti-herpetic activity of broad spectrum [13,22].

DHA (2) has exhibited strong inhibitory effects on Epstein-Barr virus early antigen (EBV-EA) activation induced by 12-O-tetradecanoylphorbol-13-acetate (TPA) and was selected to examine effects on *in vivo* two-stage mouse skin carcinogenesis. It exhibited high activity in this antitumour-promoting test [23]. Two oxidation products of methyl abietate, compounds 24a and 24b (Fig. 6), have displayed weak cytotoxicity against KB cells ($IC_{50} = 4.5$ and 5.8 μ g/ mL, respectively) [24]. Rao et al. reported a cytotoxicity study on liver cancer cells (SMMC7721) of a number of N,N'-substituted ureas (e. g. 25a-d) (Fig. 6) synthesised from DHA (2) [25]. Their IC₅₀ values were between 8.8 and 14.2 µM. The change of N'-substituted groups resulted little difference to the cytotoxicity activities of ureas, which indicated that the cytotoxicity of this kind of ureas depend strongly on the tricyclic hydrophenanthrene structure. Later on, these authors described the synthesis and antitumour activities of several acylthioureas (e. g. 26) (Fig. 6) on liver cancer cells (SMMC7721) and lung cancer cells (A549) [26]. Compound 26 exhibited higher inhibition ratio against A549 than SMMC7721 (82.89% and 61.72%, respectively). Its IC₅₀ was 6.44 μ M for A549 and 6.84 µM for SMMC7721. Recently, Zhang et al. continued this research on thioureas and studied novel α -aminophosphonates dehydroabietic acid derivatives (e. g. 27a-c) (Fig. 6) against several human cancer cell lines, including NCI-H460 (lung), A549 (lung), HepG2 (liver) and SKOV3 (ovarian) [27]. Compound 27a exhibited the best antitumour activity on NCI-H460 with IC_{50} of 3.33 μM and demonstrated better cytotoxic inhibition than 5-fluorouracil (5-FU) in all tested cell lines. Its apoptosis-inducing activity on NCI-H460 cells was studied and the results revealed that this compound showed clear cell apoptosis inducing effects. Also, C7functionalised derivative 27c exhibited high level of antitumour activities against the tested cancer cell lines and demonstrated to be more potent compared with the commercial anticancer drug 5-FU. This compound induced cell apoptosis in A549 cells. The cytotoxic activities of several amino acid-conjugate derivatives of DHA (2), compounds **28a**–**h** (Fig. 6), against four human cancer cell lines (HL60, leukaemia), (A549, lung), (AZ521, stomach) and (SK-BR-3, breast) have been reported [28]. All compounds showed cytotoxicity against HL60 cells with IC_{50} values in the range of 2.3–37.3 $\mu M.$ In addition, most of the derivatives exhibited moderate cytotoxicity against the other cancer cell lines. Among the derivatives, l-tyrosine derivative 28g exhibited potent activities against the four cancer cell lines with IC₅₀ values of 2.3 (HL60), 7.1 (A549), 3.9 (AZ521), and



Fig. 6. Antitumour DHA derivatives 24-31.

8.1 µM (SK-BR-3), respectively. Furthermore, all derivatives were shown to possess high selective cytotoxic activities for leukaemia cells, since they exhibited only weak cytotoxicity against normal lymphocyte cell line RPMI1788. In connection with this research, Zhang et al. described the synthesis and antitumour activities of novel dipeptide derivatives, compounds 29a-s (Fig. 6), derived from DHA (2) [29]. The antitumour activity screening indicated that many compounds showed moderate to high levels of inhibition activities against NCI-H460 (lung), HepG2 (liver), SKOV3 (ovarian), BEL-7404 (liver), HeLa (cervical) and HCT-116 (colon) cancer cell lines and that some displayed more potent inhibitory activities than commercial anticancer drug 5-FU. The mechanism of representative compound 29b was studied which demonstrated that it induced apoptosis in HeLa cells through a mitochondrial pathway. Recently, Pertino et al. reported on the synthesis and antiproliferative activity of some novel triazole derivatives of DHA (2), compounds **30a**–**d** and **31a**–**l** (Fig. 6) [30]. The best results were obtained for compound 31a which presented an IC₅₀ value of 6.1 µM in the SK-MES-1 cell line derived from metastatic site. Under the same experimental conditions, the IC₅₀ value of etoposide was 1.83 µM.

The redox properties of several diarylamines (*e. g.* **32a**–**d**) (Fig. 7) derived from DHA (**2**) have been investigated by cyclic



Fig. 7. Antioxidant DHA derivatives **32–33** and sedative and antipyretic DHA derivative **34**.

voltammetry, and their free radical scavenging activity was tested by reduction of the 2,2-diphenyl-1-picrylhydrazyl (DPPH) radical [31]. Compounds 32a, 32c and 32d with lower oxidation potential proved to be as active as isopropyldiphenylamine (IPPD) and superior to tert-butylhydroxytoluene (BHT). In order to probe the antioxidant effect of a series of methyl 12-imino-13-nitro dehydrodeisopropylabietate derivatives (**33a–d**) (Fig. 7), the metal ion binding abilities on Cu^{2+} and Fe^{2+} of these derivatives were studied using fluorescence quenching method [32]. The results indicated that each compound showed obvious chelation activity with the binding constants (K_A) of the 10² L mol⁻¹ order of magnitude, which implied its potential pharmacology application as antioxidant by the inhibition of Fenton reaction through chelation with Cu²⁺ and Fe²⁺. Compound **33b** has the strongest chelation activity with Cu^{2+} , while **33d** has the strongest chelation activity with Fe^{2+} , respectively.

The biological activity of a series of amino derivatives of DHA (**2**) (*e. g.* **34**) (Fig. 7) has been studied. Compound **34** produces a calming effect, anxiolytic activity, and antipyretic action comparable with that of the reference drug analgin in mice [33].

In order to explore novel scaffolds for large-conductance calcium-activated K⁺ channel openers (BK channels), Ohwada et al. selected the aromatic ring of DHA (2) for structural modification. Well-characterised BK channel openers could be used to treat acute stroke, epilepsy, and bladder overactivity. There is some evidence for the utility of BK channel openers in the treatment of asthma, hypertension, gastric hypermotility and psychoses. Thus, this research group has investigated a number of derivatives, compounds 35–38 (Fig. 8), as BK channel openers. For example, the early studies found that 12.14-dichlorodehydroabietic acid (35) showed a remarkable increase of the BK channel-opening activities [34]. Later on, the discovery of the hexahydrodibenzazepinone moiety with BK channel-opening activity was reported [35]. In this study, it was concluded that substitution with a phenyl-bearing alkynyl group on the lactam amide, compound 36, was critical for activity. A series of ureas and thioureas (e. g. 37) were also synthesised and studied as BK channel openers. Compound 37 increased the ionic current by 240% of control current at a concentration of 30 μ M [36]. Studies to improve the activity of lead compound 35 were also carried out. Thus, it was discovered that oxime ether derivatives (e. g. 38) of the benzylic ketone of 12,14dichlorodehydroabietic acid (35) were potent and effective BK channel openers [37,38]. Compound 38 was approximately twice as potent as the standard compound NS1619 and its mechanism of action has been studied [39].



Fig. 8. BK channel openers based on DHA derivatives **35–38** and anti-inflammatory derivatives **39** and **40**.

The study of Li and McChesney in 1992 described the synthesis and potential anti-inflammatory activity of DHA (**2**) derivatives. In this work, compounds **39** and **40** (Fig. 8) demonstrated weak anti-inflammatory activity [40].

2.2. Dehydroabietylamine derivatives

Dehydroabietylamine (**5**, DHAA) (Fig. 1), also called leelamine, is a synthetic primary amine having a tricyclic structure which is obtained as a part of a mixture of amines prepared from rosin. This compound and its derivatives are widely used in the fields of papermaking, and chemical industries due to its properties as resolving agent, surfactant, dye and pharmaceutical [41]. DHAA (**5**) and some



Fig. 9. Antimicrobial DHAA derivatives 41–46.

derivatives have been patented as antibacterials and anticancer agents [42,43]. One of the early studies on antimicrobial activity of DHAA (5) found that several peptoid derivatives (e. g. 41-42) (Fig. 9) were also active in the range $3-12 \mu g/mL$ against a panel of Gram-positive and Gram-negative bacteria, which included isolates which were resistant to known antibiotics [44]. The antimicrobial activity of these compounds was generally slightly more potent against Gram-positive than Gram-negative isolates. The activity against S. aureus was rapid, bactericidal, and independent of protein synthesis. Compound 42a protected S. aureus-infected mice in a simple infection model. The antibacterial activity of DHAA (5) against B. subtilis and S. aureus as well as two fungi strains (Poria placenta and Rhizopus oryzae) has also been reported [45]. DHAA (5) showed strong activity against the two bacteria, and presented fairly similar antibiotic activity against fungus of R. oryzae but little activity against fungus of P. placenta. Several quaternary ammonium salts (43-44) (Fig. 9) obtained from DHAA (5) displayed antibacterial activity against S. aureus and E. coli with MIC values in the range of 7.81–31.25 µg/mL for S. aureus, and 250–500 µg/mL for E. coli [46]. It was also found that the ammonium salts with N,Ndimethyl groups had stronger activity than those with N,Ndiethyloxyl groups. Other derivatives of DHAA (5), such as schiff base derivatives (**45a**–**h**) (Fig. 9), obtained by reaction of the amino group have shown antibacterial activity against S. aureus, B. subtilis and E. coli [47]. All the compounds exhibited bactericidal activity. Compound **45c** derived from Cl-substituted benzaldehyde was the most active towards *B. subtilis*, and compound **45d** derived from fluorinated benzaldehvde, toward S. aureus. The most active agent toward E. coli was compound 45a derived from unsubstituted benzaldehyde. The bactericidal activity of ring B derivatives of DHAA (5) such as 12-nitro-dehydroabietylamine (46a) and 12,14dinitro-dehydroabietylamine (46b) has also been reported [48]. Both compounds had strong bactericidal activity against E. coli, S. aureus and P. fluorescens.

Recently, DHAA (5) has been studied for treating melanoma showing a novel mechanism of action. It was 4.5-fold more effective at inhibiting cultured melanoma cell survival than normal cells, with average IC₅₀ values of 2 and 9.3 µM, respectively [49]. It also inhibited the growth of preexisting xenografted melanoma tumours by an average of 60% without affecting animal body weight or blood markers of major organ function. The potent biological profile of DHAA (5) has spurred the synthesis of a number of derivatives (47-56) (Fig. 10) for studying their antitumour properties. For example, a series of novel α -aminophosphonate derivatives, compounds 47a-j (Fig. 10), have been synthesised by Rao et al. and evaluated for their antitumour activities against SMMC7721 liver cancer cells [50]. Compounds 47d and 47f exhibited higher activities even at very low concentrations, and the inhibition ratios reached 75% and 79% at 0.1 uM, respectively. The inhibition ratio of compound 47i reached 99% after 72 h incubation. The derivatives with a fluorine atom and a nitro group fused to the benzene ring exhibited higher activities. The same authors have reported that dehydroabietylamine derivative 48a (TBIDOM) (Fig. 10) exhibited significant antiproliferative effects by induction of apoptosis in SMMC7721 liver cancer cells. Its mechanism may be related to the decrease in the expression of anti-apoptotic protein, Bcl-2, accompanied by a drop in the mitochondrial membrane potential and the activation of caspase-3, that led to apoptotic body formation and finally apoptosis [51]. Similarly, dehydroabietylamineflurobenzaldehyde derivative 48b (Fig. 10) displayed a significant inhibitory effect on the growth of SMMC7721 cells in a dose- and time-dependent manner (IC₅₀ = 44.5 μ M) [52]. Also, compound 48b could significantly reduce tumour weight in the H22 solid tumour mouse model in vivo. A series of Schiff bases, compounds 49a-e (Fig. 10), derived from 12-amino-N-acetyldehydroabietylamine



Fig. 10. Antitumour DHAA derivatives 47-56.

have been synthesised and evaluated for their antitumour activities on Hey-1B ovarian cancer cell line [53]. The IC₅₀ values were in the range of 15–20 μ g/mL. The urea derivative **50** (Fig. 10) showed a significant dose-dependent and time-dependent inhibition of growth of human hepatoma HepG2 cells ($IC_{50} = 3.81 \mu M$) [54]. It possibly has its effect by inhibition of glucolysis and glucolysisdependent ATP depletion. The isothiocyanate derivative 51 (Fig. 10) at concentrations ca. 1 µM displayed dose-dependent and selective cytotoxicity to endometrial cancer cells in comparison to other cancer cell lines [55]. Lin et al. have prepared several benzamide derivatives, compounds **52a**–**c** and **53a**–**c** (Fig. 10), with a carbonyl group at C-7 on ring B or a nitro group at C-12 on ring C and evaluated the influence of these groups on the cytotoxicity against PC-3 (prostate) and Hey-1B (ovarian) cancer cell lines [56]. The results showed that the presence of either a carbonyl or a nitro group increased the cytotoxic activity. Among these compounds, 52a and 53a-c showed good cytotoxicities against PC-3 cancer cells $(IC_{50} = 5.7 - 8.2 \,\mu g/mL)$ and Hev-1B cancer cells $(IC_{50} = 11.3 - 16.0 \,\mu g/mL)$ mL). These researchers also carried out a screening of 73 dehydroabietylamine derivatives as potential candidate inhibitors of the growth of liver cancer cells. In this study, N-benzoyl-12-

nitrodehydroabietylamine-7-one (54) (Fig. 10) demonstrated to have significant growth inhibitory activity in the human liver cancer cell line, HepG2 ($IC_{50} = 67.86 \ \mu g/mL$) [57]. Further research confirmed that this compound effectively induced apoptosis and inhibited HepG2 cell proliferation by blocking DNA synthesis. However, another study of this research group showed that one or two nitro groups in ring C of DHAA (5) together with aromatic imines gave highly cytotoxic compounds to normal human hepatocyte (L02) cells [58]. Recently, Rao et al. screened a series of imines, amides and ureas with a dehydroabietyl skeleton for their antitumour activities against SMMC7721 (liver), A549 (lung), C6 (glioma) and MCF-7 (breast) cancer cell lines [59]. Imines 55a-d (Fig. 10) possess noticeable antitumour activity against SMMC7721, A549, C6 and MCF-7 cancer cells, with lowest IC₅₀ values of 6.65, 0.75, 0.81, and 10.65 µM, respectively. The antitumour activity of DHAA (5) against two bladder cancer cell lines (EI and 5637), one prostate cancer cell line (PC-3), one cervical cancer cell line (HeLa) and one human T-cell leukaemia cancer cell line (Jurkat) was improved with the attachment of quinidine via thiourea bond [60]. Thus, compound 56 (Fig. 10) exhibited broad spectrum of activity $(IC_{50} = 2.07 - 3.21 \mu M)$ against the tested cancer cell lines with little toxicity to normal cells, and induced apoptosis mainly through mitochondrial-dependent pathway.

But the biological properties of DHAA (**5**) derivatives go beyond antimicrobial and antitumour activities. For example, the antiinflammatory activity of four derivatives of DHAA (**5**), compounds **57–60** (Fig. 11), has also been described. These compounds were found to be active as topical inflammation inhibitors as demonstrated in the tetradecanoylphorbol acetate induced ear oedema assay [61].

The derivative N-benzoyl-dehydroabietylamine-7-one (**61**) (Fig. 11) has been found to possess certain androgen receptor binding activity with IC₅₀ value of 83.8 μ M [62]. And the imine derivative **62** (Fig. 11) displayed tyrosine kinase inhibitory activity (FGFR1, IC₅₀ = 1.20 μ M) [63]. The antioxidative activity of derivative **63** (Fig. 11), which resulted from merging gallic acid and DHAA (**5**) through an amide bond, has also been reported [64]. Compound **63**



Fig. 11. Other bioactive DHAA derivatives 57–63.

was tested for the ability on scavenging superoxide anion radical and 1,1-diphenyl-2-picrylhydrazyl (DPPH) radical. The results indicate that the inhibitory rate of superoxide anion radical by **63** was 38.18% which is twice as much as that of L-ascorbic acid (18.35%) and its capacity on scavenging DPPH radical (IC₅₀ = 2 mg/ L) is much better than L-ascorbic acid (IC₅₀ = 236 mg/L).

In addition, DHAA (**5**) is a known inhibitor of pyruvate dehydrogenase kinase (PDK) which could have utility as a therapy for diabetes, ischaemia, lactic acidosis and cardiac insufficiency [65].

2.3. Ferruginol derivatives

Ferruginol (3) (Fig. 1) is an abietane diterpenoid that occurs in plants belonging to the Podocarpaceae, Cupressaceae, Lamiaceae and Verbenaceae families among others. This abietane has attracted much attention since it has exhibited promising bioactivities, such as antifungal and antimicrobial [66], cardioactive [67], antioxidative [68], antileishmanial and nematicidal [69]. In addition, ferruginol (3) has recently shown antitumour activity against prostate cancer [70], cytotoxicity against human pancreatic tumour cell lines [71], as well as anti-inflammatory activity [72]. This diterpene has also shown a strong protective effect in animal gastric ulcer models [73]. However, it has demonstrated high cytotoxicity which has led to the study of semisynthetic derivatives with lower toxicity. For example, Rodriguez et al. prepared a series of semisynthetic derivatives of ferruginol (3), compounds 64a-i (Fig. 12), and assessed their gastroprotective effects in the HCl/ ethanol-induced gastric lesion model in mice, as well as cytotoxicity in human gastric adenocarcinoma (AGS) and human lung fibroblasts (MRC-5) cells [74]. At 20 mg/kg, the greatest gastroprotective effects were provided by compounds 64a, 64e, **64f**, **64h**, and **64i**, all of which were as active as the reference drug lansoprazole at 20 mg/kg, reducing gastric lesions by 69, 76, 67, 72 and 61%, respectively. Compounds that showed the greatest cytotoxicity towards AGS cells were ferruginol (3), the corresponding formate 64b, acetate 64c, propionate 64d, 64e, 64f, 64g, and 64i $(IC_{50} = 18-44 \mu M)$. Ferruginol (3) and compounds 64b-g, and 64i were the most toxic compounds against fibroblasts (IC_{50}\,{=}\,19{-}56\,\mu M), with a correlation to AGS cells. The best activity/ cytotoxicity ratio was found for compound **64h**, with a lesion index comparable with lansoprazole at 20 mg/kg and cytotoxicity >1000 µM towards MRC-5 and AGS cells, respectively. Later on, this research group described the synthesis and evaluation of several new semisynthetic ester derivatives of ferruginol (3), compounds 65a-g (Fig. 12), using the same gastric model in mice and cell lines



Fig. 12. Antiulcer ferruginol derivatives 64 and 65.

[75]. Thus, the best gastroprotective effect was elicited by ferruginyl nicotinate (**65e**), reducing the lesion index by 71%, while the derivatives ferruginyl palmitate (**65a**), ferruginyl oleate (**65b**), ferruginyl 3,5-dinitrobenzoate (**65c**), ferruginyl 3-methylbenzofuran-2-carbonyl ester (**65d**), ferruginyl indoleacetate (**65f**) and ferruginyl indolebutyrate (**65g**) reduced the lesions by 50–66%. The most promising compounds were **65c**, **65e** and **65f**, presenting a gastroprotective effect higher or similar to that of ferruginol (**3**) but with a high selectivity towards the tumour AGS cells. Among these three products, the most selective towards AGS cells was **65f**, followed by **65e**, and **65c** (IC₅₀ values of 12, 22, and 29 μ M, respectively). The cytotoxicity of the above cited compounds towards fibroblasts was >1000 μ M.

The antitumour activity of sugiol (**66**) (Fig. 13), a ferruginol related diterpenoid, and several β -amino alcohol analogues, compounds **67a–d** (Fig. 13), has been reported [76]. In this work, these compounds were evaluated against three human solid tumour cell lines A2780 (ovarian), SW1573 (lung) and WiDr (colon) and showed more potent activities (IC₅₀ values in the range 1.5–6.7 μ M) than the parent compound **66** (IC₅₀ values in the range 23–>50 μ M).

Gigante et al. prepared four catechols, compounds **68a–d** (Fig. 13) from abietic acid and evaluated several biological activities, including antifungal, antitumour, antiviral and inhibition of nitric oxide (NO) production [77]. Compounds **68a** and **68d** exhibited potent antifungal activity against dermatophytes *Microsporum canis, Trichophyton mentagrophytes* and *Epidermophyton floccosum*, being more potent (MIC = 13.1–210.0 μ M) than the control drug fluconazole (MIC = 52.2–208.9 μ M). Catechols **68a–d** were potent inhibitors of NO production (IC₅₀ = 5.2–18.5 μ M) and also displayed moderate antitumour activity. Compound **68a** was found to have antiherpetic activity and moderate anti-HIV activity (EC₅₀ = 275 μ M). Later on, this research group demonstrated the potential application of compound **68a** as antioxidant [78].



Fig. 13. Bioactive ferruginol derivatives 66-73.

Tada et al. developed a synthetic route to (+)-ferruginol (**3**) and the corresponding enantiomer (–)-ferruginol. Using this methodology, this research group prepared various oxidised abietanes, including catechols **69a** and **69b** (Fig. 13), 6-oxoferruginol (**70a**) and 6β -hydroxyferruginol (**70b**) (Fig. 13) and evaluated their antimicrobial activity against methicillin-resistant *S. aureus* (MRSA) and vancomycin-resistant *Enterococcus* (VRE) [79]. Compounds **69–70** showed potent activity with MIC values of 4–8 µg/mL against MRSA and 4–16 µg/mL against VRE. Non-natural (–)-ferruginol showed stronger activity than natural (+)-ferruginol (**3**).

Recently, Sánchez-Moreno et al. reported on the antileishmanial activity (*Leishmania infantum* and *Leishmania braziliensis*) of several phenol abietanes related to ferruginol (**3**), compounds **71–73** (Fig. 13) [80]. These compounds were more active ($IC_{50} = 14.0-31.6 \mu M$) and less toxic than the reference drug glucantime.

2.4. Carnosol and carnosic acid derivatives

Carnosol (CS, **74**) (Fig. 14) and carnosic acid (CA, **4**) (Fig. 1) are found in the popular Lamiaceae herbs, sage (*Salvia officinalis*) and rosemary (*Rosmarinus officinalis*). Both possess an o-diphenol structure (catechol) which undergoes oxidation easily and provides potent antioxidant activity. The early studies on biological activity of derivatives of these abietanes considered particularly the antioxidant activity. For example, Nakatani et al. reported on the antioxidant activity of compounds **74b–d**, **75a–b** and **76b–e** (Fig. 14) derived from carnosol (**74**) and rosmanol (**76a**), a related catechol, respectively [81]. In general, when the phenol is fully substituted there is a decrease in activity while mono-substituted derivatives were as strong antioxidants as the parent compounds. Preparative-scale incubations of CA (**4**) with *Nocardia* sp. led to the isolation of derivatives **77** and **78** (Fig. 14), which were evaluated for antioxidant activity using the DPPH free-radical scavenging assay



Fig. 14. Bioactive carnosic acid derivatives 74–83.

[82]. Both compounds showed activities similar to that of tocopherol and CA (**4**).

Takeya et al. prepared several semisynthetic derivatives from CA (4), including compounds **79a–b** (Fig. 14), and examined the cytotoxic activity using P388 murine leukaemia cells [83]. Substitution of the phenol groups decreased the cytotoxicity, compound **79b** (IC₅₀ = 5.9 µg/mL), but the free hydroxyl groups along with the ester group at C-20, compound **79a** (IC₅₀ = 0.6 µg/mL), gave more potent activity than CA (4) (IC₅₀ = 1.2 µg/mL). In another study, compound **79b** also showed cytotoxicity against breast (MCF-7) cancer cell line (IC₅₀ = 69 µM) as well as activity against *Mycobacterium tuberculosis* (MIC = 28 µM), being more potent than carnosol (**74**) [84].

The gastroprotective activity of carnosic acid (4) derivatives has also been reported. For example, Schmeda-Hirschmann et al. described the synthesis and evaluation of a series of γ -lactone derivatives (e. g. 80a–d) (Fig. 14) of carnosic acid (4) in the HCl/ ethanol-induced gastric lesion model in mice, as well as cytotoxicity in human gastric adenocarcinoma (AGS) and human lung fibroblasts (MRC-5) cells [85]. Compounds 80a-d showed the highest gastroprotective effect (65-82% lesion reduction), however, compound **80a** was more cytotoxic ($IC_{50} = 130 \ \mu M$ for MRC-5 and 103 µM for AGS) than compounds 80c and 80d, which presented IC₅₀ values of >1000 μ M against MRC-5 cells and >1000 and 822 µM toward AGS cells, respectively. Later on, this research group reported on the gastroprotective effect of further derivatives of carnosic acid (4), compounds 81-83 (Fig. 14). At 10 mg/kg, compounds 81–83 were more effective preventing gastric lesions (73–78% lesion reduction) than the reference drug lansoprazole at the same dose (64% lesion reduction). These compounds showed the best gastroprotective effect combines with the lowest cytotoxicity [86]. Their mechanism of action has also been studied, which includes protection against cell damage induced by sodium taurocholate, increase in glutathione content, stimulation of prostaglandin E₂ synthesis and cell proliferation [87].

3. Conclusion

A number of derivatives of natural aromatic abietanes have been synthesised and studied for biological properties. The encouraging findings warrant further interesting studies in this area of research, which could provide with novel chemotherapeutics and chemical probes.

Acknowledgements

Financial support from the Universidad de Valencia is gratefully acknowledged.

References

- [1] D.J. Newman, G.M. Cragg, J. Nat. Prod. 75 (2012) 311-335.
- [2] J.R. Hanson, Nat. Prod. Rep. 30 (2013) 1346–1356 and preceding contributions of the author.
- [3] A. San Feliciano, M. Gordaliza, M.A. Salinero, J.M. Miguel del Corral, Planta Med. 59 (1993) 485–490.
- [4] L.F. Fieser, W.P. Campbell, J. Am. Chem. Soc. 60 (1938) 159–170.
- [5] C.W. Brandt, L.G. Neubauer, J. Chem. Soc. (1939) 1031–1037.
- [6] (a) G.C. Harris, J. Am. Chem. Soc. 70 (1948) 3671–3674;
 (b) T. Norin, B. Winell, Acta Chem. Scand. 26 (1972) 2289–2296;
 - (c) A. Ulubelen, M. Miski, T.J. Mabry, J. Nat. Prod. 44 (1981) 119–124.
- [7] N. Okamura, Y. Fujimoto, S. Kuwabara, A. Yagi, J. Chromatogr. A 679 (1994) 381–386.
- [8] H. Wada, S.-I. Kodato, M. Kawamori, T. Morikawa, H. Nakai, M. Takeda, S. Saito, Y. Onoda, H. Tamaki, Chem. Pharm. Bull. 33 (1985) 1472–1487.
- 9] B. Sepúlveda, L. Astudillo, J.A. Rodríguez, T. Yañez, C. Theoduloz, G. Schmeda-Hirschmann, Pharmacol. Res. 52 (2005) 429–437.
- [10] S. Savluchinske-Feio, M.J. Marcelo Curto, B. Gigante, J.C. Roseiro, Appl. Microbiol. Biotechnol. 72 (2006) 430–436.

- [11] S. Savluchinske-Feio, B. Gigante, J.C. Roseiro, M.J. Marcelo Curto, J. Microbiol. Methods 35 (1999) 201-206.
- [12] B. Gigante, A.M. Silva, M.J. Marcelo Curto, S. Savluchinske-Feio, J.C. Roseiro, L.V. Reis, Planta Med. 68 (2002) 680-684.
- [13] M.A. González, D. Pérez-Guaita, J. Correa-Royero, B. Zapata, L. Agudelo, A. Mesa-Arango, L. Betancur-Galvis, Eur. J. Med. Chem. 45 (2010) 811-816.
- [14] B. Zapata, M. Rojas, L. Betancur-Galvis, A.C. Mesa-Arango, D. Pérez-Guaita, M.A. González, Med. Chem. Comm. 4 (2013) 1239–1246.
- [15] X.-T. Xu, X.-L. Ma, W.-G. Duan, L. Chen, B. Cen, Q.-J. Mo, J.-Y. Wang, Holzforschung 67 (2013) 107–112.
- [16] G.-F. Su, L.-N. Huo, R. Chen, F.-L. Zhao, Y.-F. Guan, Acta Chim, Sin, 66 (2008) 2717-2724.
- [17] W. Gu, S. Wang, Eur. J. Med. Chem. 45 (2010) 4692–4696.
 [18] W. Gu, C. Qiao, S.-F. Wang, Y. Hao, T.-T. Miao, Bioorg. Med. Chem. Lett. 24 (2014) 328-331.
- [19] W. Gu, R. Wu, S. Oi, C. Gu, F. Si, Z. Chen, Molecules 17 (2012) 4634–4650.
- [20] T. Fonseca, B. Gigante, M.M. Marques, T.L. Gilchrist, E. De Clercq, Bioorg. Med. Chem. 12 (2004) 103-112.
- [21] J.R. Tagat, D.V. Nazareno, M.S. Puar, S.W. McCombie, A.K. Ganguly, Bioorg. Med. Chem. Lett. 4 (1994) 1101–1104.
- [22] L.S. Agudelo-Gómez, L.A. Betancur-Galvis, M.A. González, Pharmacologyonline 1 (2012) 36-42.
- [23] R. Tanaka, H. Tokuda, Y. Ezaki, Phytomedicine 15 (2008) 985–992.
- [24] S. Prinz, U. Müllner, J. Heilmann, K. Winkelmann, O. Sticher, E. Haslinger, A. Hüfner, J. Nat. Prod. 65 (2002) 1530-1534.
- [25] X. Rao, Z. Song, L. He, W. Jia, Chem. Pharm. Bull. 56 (2008) 1575-1578.
- [26] X.-P. Rao, Y. Wu, Z.-Q. Song, S.-B. Shang, Z.-D. Wang, Med. Chem. Res. 20 (2011) 333-338.
- [27] (a) X.-C. Huang, M. Wang, Y.-M. Pan, G.-Y. Yao, H.-S. Wang, X.-Y. Tian, J.-K. Qin, Y. Zhang, Eur. J. Med. Chem. 69 (2013) 508-520; (b) X.-C. Huang, M. Wang, Y.-M. Pan, X.-Y. Tian, H.-S. Wang, Y. Zhang, Bioorg.
- Med. Chem. Lett. 23 (2013) 5283-5289. [28] M. Ukiya, T. Kawaguchi, K. Ishii, E. Ogihara, Y. Tachi, M. Kurita, Y. Ezaki,
- M. Fukatsu, Y. Kushi, T. Akihisa, Chem. Biodiv. 10 (2013) 1260-1268.
- [29] X.-C. Huang, M. Wang, H.-S. Wang, Z.-F. Chen, Y. Zhang, Y.-M. Pan, Bioorg. Med. Chem. Lett. 24 (2014) 1511-1518.
- [30] M.W. Pertino, V. Verdugo, C. Theoduloz, G. Schmeda-Hirschmann, Molecules 19 (2014) 2523-2535.
- [31] M.A. Esteves, N. Narender, M.J. Marcelo-Curto, B. Gigante, J. Nat. Prod. 64 (2001) 761-766.
- [32] K. Wang, X. Liu, X. Yi, H. Wang, Y. Zhang, Y. Pan, Med. Chem. Res. 21 (2012) 2494 - 2500.
- [33] I.A. Tolmacheva, A.V. Tarantin, A.A. Boteva, L.V. Anikina, Y.B. Vikharev, V.V. Grishko, A.G. Tolstikov, Pharm. Chem. J. 40 (2006) 489-493.
- [34] T. Ohwada, T. Nonomura, K. Maki, K. Sakamoto, S. Ohya, K. Muraki, Y. Imaizumi, Bioorg. Med. Chem. Lett. 13 (2003) 3971-3974.
- [35] T. Tashima, Y. Toriumi, Y. Mochizuki, T. Nonomura, S. Nagaoka, K. Furukawa, H. Tsuru, S. Adachi-Akahane, T. Ohwada, Bioorg. Med. Chem. 14 (2006) 8014-8031.
- [36] Y.-M. Cui, E. Yasutomi, Y. Otani, T. Yoshinaga, K. Ido, K. Sawada, T. Ohwada, Bioorg. Med. Chem. Lett. 18 (2008) 5201-5205.
- [37] Y.-M. Cui, E. Yasutomi, Y. Otani, T. Yoshinaga, K. Ido, K. Sawada, M. Kawahata, K. Yamaguchi, T. Ohwada, Bioorg. Med. Chem. Lett. 18 (2008) 6386–6389.
- [38] Y.-M. Cui, E. Yasutomi, Y. Otani, K. Ido, T. Yoshinaga, K. Sawada, T. Ohwada, Bioorg. Med. Chem. 18 (2010) 8642-8659.
- [39] G. Gessner, Y.-M. Cui, Y. Otani, T. Ohwada, M. Soom, T. Hoshi, S.H. Heinemann, PNAS 109 (2012) 3552-3557.
- [40] W.-S. Li, J.D. McChesney, J. Pharm. Sci. 81 (1992) 646-651.
- [41] X. Rao, Z. Song, H. Gao, Chem. Bull. 69 (2006) 168-172.
- [42] P.C. De Vissier, G.J. Platenburg, J.T. Van Dissel, WO 2008016300 A2, 2008.
- [43] G.P. Robertson, C.D. Raghavendragowda, S.V. Madhunapantula, O.F. Kuzu, G.S. Inamdar, US Patent 20120141578 A1, 2012.
- [44] B. Goodson, A. Ehrhardt, S. Ng, J. Nuss, K. Johnson, M. Giedlin, R. Yamamoto, W.H. Moos, A. Krebber, M. Ladner, M.B. Giacona, C. Vitt, J. Winter, Antimicrob. Agents Chemother. 43 (1999) 1429–1434.
- [45] Y. Li, Chin. J. Struct. Chem. 27 (2008) 1089-1092.
- [46] Y. Wang, C. Yang, Y. Zhou, Z. Song, Nat. Prod. Res. Dev. 10 (1998) 59-63. [47] X. Rao, Z. Song, A.B. Radbil', B.A. Radbil', Russ. J. Appl. Chem. 80 (2007) 1373-1375.
- [48] S. Zhang, Z. Lin, J. Chem. Ind. Eng. (China) 60 (2009) 3077-3081.
- [49] R. Gowda, S.V. Madhunapantula, O.F. Kuzu, A. Sharma, G.P. Robertson, Mol. Cancer Ther. 13 (2014) 1679-1689.
- [50] X. Rao, Z. Song, L. He, Heteroat. Chem. 19 (2008) 512-516.

- [51] F. Li, L. He, Z. Song, J.-C. Yao, X.-P. Rao, H.-T. Li, J. Pharm. Pharmacol. 60 (2008) 205-211
- [52] L. Liu, F. Li, L. He, X.-P. Rao, Z.-Q. Song, Chin. Pharm. J. 48 (2013) 269-274.
- [53] S. Zhang, Z. Lin, Chin. J. Pharm. 41 (2010) 566-579.
- [54] J.X. Xie, L. He, L.-Y. Zhang, X.-P. Rao, Z.-Q. Song, J. China Pharm. Univ. 41 (2010) 160-165.
- [55] T.C. Horan, M.A. Zompa, C.T. Seto, K.K. Kim, R.G. Moore, T.S. Lange, Investig. New Drugs 30 (2012) 1460–1470.
- [56] Y. Chen, Z.-X. Lin, A.-M. Zhou, Nat. Prod. Res. 26 (2012) 2188-2195.
- [57] L.-Y. Lin, Y.-L. Bao, Y. Chen, L.-G. Sun, X.-G. Yang, B. Liu, Z.-X. Lin, Y.-W. Zhang,
- C.-L. Yu, Y. Wu, Y.-X. Li, Chem. Biol. Interact, 199 (2012) 63-73. [58] C.-X. Liu, Z.-X. Lin, X. Yu, Z. Lu, A.-M. Zhou, Y.-L. Bao, Pharmazie 68 (2013)
- 861-865. [59] X. Rao, X. Huang, L. He, J. Song, Z. Song, S. Shang, Comb. Chem. High Throughput Screen. 15 (2012) 840–844.
- [60] Y. Xing, W. Zhang, J. Song, Y. Zhang, X. Jiang, R. Wang, Bioorg. Med. Chem. Lett. 23 (2013) 3868-3872
- [61] W.W. Wilkerson, W. Galbraith, I. DeLucca, R.R. Harris, Bioorg. Med. Chem. Lett. 3 (1993) 2087–2092.
- [62] Y. Chen, Z.-X. Lin, Chem. Ind. For. Prod. 29 (2009) 121–124.
- [63] T.-T. Zhou, L. He, M. Yang, L.-Y. Zhang, J.-G. He, X.-P. Rao, Chin. J. Nat. Med. 11 (2013) 506-513.
- [64] Z. Lu, C. Liu, X. Yu, Z. Lin, Chin. J. Org. Chem. 33 (2013) 562–567.
- [65] T.D. Aicher, R.E. Damon, J. Koletar, C.C. Vinluan, L.J. Brand, J. Gao, S.S. Shetty, E.L. Kaplan, W.R. Mann, Bioorg. Med. Chem. Lett. 9 (1999) 2223-2228.
- [66] J. Becerra, C. Flores, J. Mena, P. Aqueveque, J. Alarcón, M. Bittner, V. Hernández, M. Hoeneisen, E. Ruiz, M. Silva, J. Chil. Chem. Soc. 47 (2002) 151-157.
- [67] A. Ulubelen, H. Birman, S. Oksuz, G. Topcu, U. Kolak, A. Barla, W. Voelter, Planta Med. 68 (2002) 818-821.
- [68] M. Ono, M. Yamamoto, C. Masuoka, Y. Ito, M. Yamashita, T. Nohara, J. Nat. Prod. 62 (1999) 1532-1537.
- [69] V. Samoylenko, D.C. Dunbar, M.A. Gafur, S.I. Khan, S.A. Ross, J.S. Mossa, F.S. El-Feraly, B.L. Tekwani, J. Bosselaers, I. Muhammad, Phytother. Res. 22 (2008) 1570 - 1576
- [70] M. Bispo de Jesus, W.F. Zambuzzi, R.R. Ruela de Sousa, C. Areche, A.C. Santos de Souza, H. Aoyama, G. Schmeda-Hirschmann, J.A. Rodríguez, A.R.M. de Souza Brito, M.P. Peppelenbosch, J. den Hertog, E. de Paula, C.V. Ferreira, Biochimie 90 (2008) 843-854.
- [71] M. Fronza, R. Murillo, S. Slusarczyk, M. Adams, M. Hamburger, B. Heinzmann, S. Laufer, I. Merfort, Bioorg. Med. Chem. 19 (2011) 4876-4881.
- [72] H.L. Chen, K.W. Lin, K.H. Gan, J.P. Wang, S.J. Won, C.N. Lin, Fitoterapia 82 (2011) 219-224.
- [73] J.A. Rodriguez, C. Theoduloz, T. Yañez, J. Becerra, G. Schmeda-Hirschmann, Life Sci. 78 (2006) 2503-2509.
- [74] C. Areche, J.A. Rodriguez, I. Razmilic, T. Yañez, C. Theoduloz, G. Schmeda-Hirschmann, J. Pharm. Pharmacol. 59 (2007) 289-300.
- [75] M. Espinoza, L.S. Santos, C. Theoduloz, G. Schmeda-Hirschmann, J.A. Rodriguez, Planta Med. 74 (2008) 802-808.
- [76] I. Córdova, L.G. León, F. León, L. San Andrés, J.G. Luis, J.M. Padrón, Eur. J. Med. Chem. 41 (2006) 1327-1332.
- [77] B. Gigante, C. Santos, A.M. Silva, M.J.M. Curto, M.S.J. Nascimento, E. Pinto, M. Pedro, F. Cerqueira, M.M. Pinto, M.P. Duarte, A. Laires, J. Rueff, J. Gonçalves, M.I. Pegado, M.L. Valdeira, Bioorg. Med. Chem. 11 (2003) 1631–1638.
- [78] G.C. Justino, C.F. Correia, L. Mira, R.M.B. Dos Santos, J.A.M. Simões, A.M. Silva, C. Santos, B. Gigante, J. Agric. Food Chem. 54 (2006) 342-348.
- Z. Yang, Y. Kitano, K. Chiba, N. Shibata, H. Kurokawa, Y. Doi, Y. Arakawa, [79] M. Tada, Bioorg. Med. Chem. 9 (2001) 347-356.
- [80] I. Ramírez-Macías, C. Marín, R. Chahboun, F. Olmo, I. Messouri, O. Huertas, M.J. Rosales, R. Gutierrez-Sánchez, E. Alvarez-Manzaneda, M. Sánchez-Mor-
- eno, Mem. Inst. Oswaldo Cruz 107 (2012) 370-376. R. Inatani, N. Nakatani, H. Fuwa, Agric. Biol. Chem. 47 (1983) 521-528.
- [82] M. Hosny, H.A. Johnson, A.K. Ueltschy, J.P.N. Rosazza, J. Nat. Prod. 65 (2002) 1266-1269.
- [83] Y. Aoyagi, Y. Takahashi, Y. Satake, K. Takeya, R. Aiyama, T. Matsuzaki, S. Hashimoto, T. Kurihara, Bioorg. Med. Chem. 14 (2006) 5285-5291.
- [84] A.A. Hussein, J.J.M. Meyer, M.L. Jimeno, B. Rodríguez, J. Nat. Prod. 70 (2007) 293-295.
- [85] M.W. Pertino, C. Theoduloz, J.A. Rodriguez, T. Yañez, V. Lazo, G. Schmeda-Hirschmann, J. Nat. Prod. 73 (2010) 639-643.
- [86] C. Theoduloz, M.W. Pertino, J.A. Rodriguez, G. Schmeda-Hirschmann, Planta Med. 77 (2011) 882-887.
- [87] C. Theoduloz, M.W. Pertino, G. Schmeda-Hirschmann, Molecules 19 (2014) 581-594.