- 1 Bovine paramphistomosis in Galicia (Spain): prevalence, intensity,
- 2 aetiology and geospatial distribution of the infection.

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39 Abstract

40 This study explored various aspects of the epidemiology of paramphistomosis in 41 Galicia, the main cattle producing region in Spain. A total of 589 cows from different 42 farms were selected in the slaughterhouse for examination of the forestomachs for the 43 presence of Paramphistomidae flukes. Paramphistomes were found in 111/589 cows 44 (18.8%; 95% CI: 15.7%-21.9%), with higher prevalences of infection in beef than in 45 dairy cows (29.2% vs 13.9%). Although the number of flukes per animal was generally 46 low (median=266), some cows harboured large burdens (up to 11895), which may 47 have harmful effects on their productivity. Cows with higher parasite burdens also 48 excreted greater numbers of eggs in faeces, so that heavily parasitized cows play an 49 important role in the transmission of paramphistomosis. This role is especially relevant 50 in Galicia, where the roe deer, which is the only wild ruminant in the study area, was 51 found not to be a reservoir for the infection. The use of morpho-anatomical and 52 molecular techniques provided reliable confirmation that Calicophoron daubneyi is the 53 only species of the family Paramphistomidae that parasitizes cattle in Galicia. The 54 environmental data from the farms were used in Bayesian geostatistical models to 55 predict the probability of infection by *C.daubneyi* throughout the region. The results

56 revealed the role of environmental risk factors in explaining the geographical

57 heterogeneity in the probability of infection in beef and dairy cattle. These explanatory

- 58 factors were used to construct predictive maps showing the areas with the highest risk
- 59 of infection and the uncertainty associated with the predictions.
- 60

61 Keywords

- Paramphistomosis, *Calicophoron daubneyi*, cattle, roe deer, epidemiology, Bayesian
 geostatistical model
- 64

65 Introduction

66 Paramphistomosis is a parasitic infection caused by digenetic trematodes belonging to

the family Paramphistomidae Fischoeder, 1901, which includes many genera and

68 species (e.g. Paramphistomum spp., Calicophoron spp., Cotylophoron spp.,

69 *Gigantocotyle* spp., etc.) that inhabit the gastrointestinal tract of wild and domestic

ruminants throughout the world [1]. Paramphistomes have a heteroxenous life cycle

71 that involves freshwater snails (intermediate hosts). The parasite larvae develop until

reaching a stage (cercaria) that emerges from the mollusc and typically encysts on

vegetation, hard surfaces or water, and then develops into a stage (metacercaria) that

is ingested by grazing ruminants (final host). In the ruminant host, juvenile parasites

75 first locate in the small intestine and feed on the intestinal mucosa. As they grow, the

76 parasites migrate upwards to the reticulum and rumen where they spend the remainder

of their adult lives, shedding eggs that contaminate snail habitats.

As regards the pathology caused by the paramphistomes in domestic ruminants, most authors consider that immature worms migrating in the small intestine can provoke severe damage, including death, whereas the adult flukes established in rumen and reticulum are not considered very harmful [2, 3, 4]. Nevertheless, ruminal lesions due to heavy infections by adult worms have been recorded [5, 4]. Digestion and absorption may be affected, resulting in diarrhoea, anorexia, anaemia and weakness [6, 7, 8]. The

effect of the infection on animal production remains controversial [9, 10], partly
because detrimental effects of paramphistomes can be masked by concurrent
infections with other helminths (e.g. *Fasciola hepatica* and gastrointestinal nematodes)
that usually infect grazing ruminants. Moreover, as with other parasitic infections, the
pathology of paramphistosis also depends on the immune status of infected animals,
the size of parasite burdens and the species of parasite involved in the infections [4,
11].

91 Paramphistomosis is highly prevalent in tropical and subtropical countries [2, 12, 13, 8] 92 where it causes high morbidity, mainly in animals raised by use of traditional husbandry 93 systems, in which periods of nutritional stress occur. In Europe, the presence of 94 paramphistomes in cattle had been considered virtually harmless [14]. However, 95 Dorchies et al. [3] have described some cases of serious illness caused by 96 paramphistomes. Furthermore, epidemiological studies conducted in central France 97 [15] have shown that the prevalence of infection with *Calicophoron daubneyi* [16, 17] in 98 cattle increased significantly, from 5.2% to 44.7%, between 1990 and 1999. 99 Paramhistomosis has also been detected on farms in UK and Wales [18] and Ireland 100 [19], indicating that this pathology should be considered in the differential diagnosis of 101 other enteric processes in cattle. In Spain, infections by Paramphistomum cervi have 102 been reported in cows and deer [20, 21]. Díaz et al. [22, 23] detected, by coprological 103 techniques, infections by paramphistomes in 26%-36% cattle farms in Galicia, which 104 appears to indicate a high prevalence of bovine paramphistomosis in the region. 105 However, documentation of the presence of infection and/or the raw data on its 106 prevalence, without any assessment of the spatial variability existing in such a wide 107 region, is of limited interest and does not indicate the real risk of infection in areas with 108 different climatic and environmental conditions, which may affect the life cycle of the 109 parasite. Moreover, other circumstances such as the existence of wild reservoirs for 110 paramphistomes have not been evaluated.

111 All of the above led us to carry out a comprehensive epidemiological study on cattle 112 paramphistomosis in Galicia, which included the following steps: 1) at bovine host 113 level, we determined the parasite burdens in rumens and reticula as well as the 114 distribution of the parasites within these organs. We also identified, by 115 morphoanatomical and molecular analysis, the species of Paramphistomidae worms 116 found at necropsy, and we determined the association between the amount of flukes in 117 the rumen and reticulum and the numbers of parasite eggs in faecal samples; 2) at the 118 host level, i.e. in roe deer (*Capreolus capreolus*), which is the only wild ruminant found 119 in the vicinity of cattle farms, we performed necropsies to detect the presence of 120 paramphistomes; and 3) at geospatial level, we investigated the distribution of the 121 cattle paramphistomosis, exploring the linkages between the spatial patterns and some 122 climatic and environmental factors that are thought to affect different phases of the 123 parasite life cycle (e.g. survival and distribution of the intermediate host and its 124 probability of infection, the development of the intra-mollusc larval stages and the 125 infection risk of cattle, among other factors). Moreover, we constructed predictive maps 126 to help decision-makers spatially target the monitoring and control of paramphistomosis 127 in dairy and beef cattle. All statistical analyses were performed in a Bayesian 128 framework to incorporate uncertainty about model parameters.

129

130 Materials and methods

131 Study area

The present study was carried out in Galicia (NW Spain), which is located between latitudes 41° 49' to 43° 47'N and longitudes 6° 42' to 9° 18'W, and surrounded by the Cantabria Sea to the north and the Atlantic Ocean to the west. The climate in the region is temperate maritime, with an annual average temperature of 11.6 °C and annual average rainfall of 1065 mm. The altitude ranges between 0 and 2129 m. The soil mainly consists of metamorphic and igneous rocks, which in hydrological terms represents a very heterogeneous and anisotropic environment.

139 Galicia covers a total surface area of 29575 km², administratively divided into 315 140 municipalities with very different cattle farming activity and stocking rate per surface 141 unit. According to the 2008 livestock census, there were 339530 dairy cows (99% on 142 farms in the northern half of the region), and 221917 beef cows (on farms spread over 143 a larger area extending to the South-East of the region). Grasslands occupy 144 approximately 60% of the useful agricultural land and cows usually graze throughout 145 the year, mainly on beef cattle farms. The type of livestock husbandry and the climatic 146 characteristics of the region favour grazing-linked transmission of helminthosis. 147

148 Sampling

149 A slaughterhouse that processes cattle from the whole region was visited fortnightly

150 during 2008. At each visit, 20 adult cows (over 2 years old) were selected at random to

151 determine the existence of infections by paramphistomes within the rumens and/or

152 reticula, as well as the occurrence of trematode eggs in the faeces. The rumen, reticula

and stool samples from a total of 589 cows, all from different farms (Fig. 1), were

154 removed and transported to the laboratory promptly.

155 The forestomachs from 235 roe deer killed during the 2008 hunting season throughout

156 Galicia were also collected and examined for the presence of flukes.

157

158 Parasitological techniques

159 Each forestomach was thoroughly examined for recovery and counting of flukes;

160 moreover, in 75 out of the parasitized cows, the anatomic distribution of flukes within

161 the rumen (atrium, rumenoreticular sulcus, ventral sac and dorsal sac) was also

162 recorded.

163 Species identification was carried out by conventional microscopy and subsequent

164 confirmation by molecular techniques. A sample of 50 paramphistomes (or all of those

165 present, if parasite burdens was not as high as this) was collected at random from each

166 parasitized cow and then split into 2 subsamples. The parasites in one of the

167 subsamples were preserved in 70% ethanol until being stained for microscopic

168 examination, while specimens in the other subsample were frozen individually at -85 °C

169 until extraction of the DNA for molecular analysis by PCR. In those 75 cows in which

170 the location of the parasites was recorded, samples were taken from each anatomical

171 region following the pattern described. Species were identified as follows:

172 1) A total of 618 randomly selected alcohol-fixed specimens (from different cows and

173 anatomical locations) were stained with borax carmine, mounted on permanent slides

and examined by microscopy and microphotography in order to measure the length,

175 width and area of the whole body, the oral and ventral suckers, the anterior and

176 posterior testis and the ovary. The species were identified according to the criteria

177 outlined by Dinnik, J.A. [16] and Eduardo, S.L. [17].

178 2) The species identity was further confirmed by molecular analysis of 82 individually

179 frozen parasite specimens. Samples of DNA extracted from 82 specimens were

180 analyzed as reported by Martínez-Ibeas et al. [25]. Briefly, the second internal

181 transcribed spacer (ITS-2) region of ribosomal DNA from each of the individual flukes

182 was amplified by PCR, with primers specific to the species, previously identified by

183 morphological techniques (F: 5' TGCATACTGCTTTGAACATCG 3' and R: 5'

184 GTTCAGCGGGTATTCACGTC 3'). The sequences of the amplification products were

185 compared with those available in GenbankTM for species identification.

186 Faeces were analysed by a standard sedimentation technique. Faecal samples (10 g)

187 were first subjected to 3 successive sedimentation processes, and the sediments were

188 then examined by microscopy to count the paramphistomid eggs. The analytical

189 sensitivity of this technique was 2 eggs per gram of faeces (epg).

190

191 Factors potentially associated with infections by paramphistomes

192 A total of 10 variables were tested for their potential association with bovine

193 paramphistomosis. Data on cow-related variables, such as age, type of production

194 (milk or beef) and livestock density (cows/km² in year 2008) in the municipalities where

195 source farms were located were provided by the Galician Animal Production Service. 196 Data on environmental variables (land cover, elevation, slope, type of soil and soil 197 permeability) for the geographical coordinates of originating farms were provided by the 198 Galician Cartographic Office (SITGA). The land cover data supplied corresponded to 199 the following 8 categories of the CORINE Land Cover map: 1: forest, 2: permanent 200 crops, 3: arable land, 4: pastures, 5: wetland, 6: shrub, 7: mines and 8: urban zones. 201 Data on elevation (m) and slope (%) were obtained from a 5-meter resolution Digital 202 Elevation Model (DEM), which was created by the National Geographic Institute of 203 Spain, within the National Project of Aerial Orthophotography. The source of lithological 204 data for soil classification was the 1:50000 digital geological map constructed by the 205 Geological and Mining Institute of Spain (MAGNA project). According to this map, there 206 are 10 different soil types in Galicia: 1: limestone and dolomite, 2: acid igneous rock, 3: 207 cenozoic deposits, 4: gneiss, 5: amphibolite, 6: slate and guartzite, 7: shale, 8: eclogite 208 and granulite, 9: basalt and peridotite, and 10: serpentinite. The digital geolithological 209 map was also used to create a permeability map from which soil permeability data 210 were obtained and then grouped into three categories: 1: low permeability (≤ 0.5 cm/h). 211 2: medium permeability (0.5-12.5 cm/h) and 3: high permeability (> 12.5 cm/h). 212 The climatic variables included in the study were the annual mean temperature and 213 total rainfall calculated from the data recorded at the 67 official weather stations in 214 Galicia during the period 2004-2008 (www.meteogalicia.es). The average annual values were then used to make a kriging (1 km² grid), for which data for each farm 215 216 coordinate were interpolated.

217

218 Statistical analysis

219 The prevalence of the Paramphistomidae flukes was first analyzed in terms of the type

of cattle (beef or dairy) and age (less than 6 years; between 7 and 9 years; and more

- than 9 years) using a Generalized Linear Model (GLM) (specifically a logistic
- 222 regression model, in which the response variable was a binary variable that represents

223 the presence or absence of the parasite in each cow sampled). We also used a GLM to 224 model the relationship between the positive coprology with the type of cattle and age. A 225 linear model was used to analyze the relationship between the total number of 226 paramphistomes observed in infected cows (in particular its logarithm) and the type of 227 cattle and age. A linear model was also used to analyze the relationship between the 228 number of eggs observed in infected cows (in particular, a Box-Cox transformation of 229 this variable) and the type of cattle and age. Finally, the relationship between the latter 230 two response variables was also analyzed by a Linear Model (with logarithmic 231 transformation of the parasite burden). As previously mentioned, the INLA methodology 232 was used to implement these five types of analysis within a Bayesian framework to 233 incorporate uncertainty about model parameters [24]. 234 The spatial variation in the probability of infection in beef and dairy cattle was modelled 235 by a hierarchical Bayesian spatial approach (see 26), specifically a point-reference

spatial model [27]. These models are highly suitable for situations (as in the present

study) in which data are observed at continuous locations occurring within a defined

spatial domain (geo-referenced Bernoulli data). Note that these models can also be

239 considered as a spatial extension of logistic regression models because the modelling

240 process describes the variability in the response variable as a function of the

241 explanatory variables, with the addition of a stochastic spatial effect, which models the

residual spatial autocorrelation [28].

243 More specifically, the response variable is a binary variable that represents the

presence or absence of the parasite in each cow sampled: Z_i is 1 if cow *i* is infected

and 0 if not. Consequently, the conditional distribution of the data is $Z_i \sim Ber(\pi_i)$,

assuming that observations are conditionally independent given π_i , which is the

probability of occurrence at location i (I = 1,...,n).

At the first stage of the hierarchical model, we modelled the occurrence of parasite as a GLM by using the usual (for binary data) logit link function, but incorporating a spatial effect. That is,

 $logit(\pi_i) = X_i\beta + W_i$

251 where β represents the vector of the regression coefficients, X is the matrix of 252 covariates, W represents spatial random effects and the logit transformation is defined as $logit(\pi_i) = \log(\frac{\pi_i}{1-\pi_i})$. 253 254 The second stage of the hierarchical model is used to incorporate the uncertainty about 255 the parameters (before taking into account the observations) used in the first level. 256 Following Bayesian reasoning, the parameters are treated as random variables, and 257 prior knowledge is incorporated via prior distributions. In particular, for the parameters 258 involved in the fixed effects, we use Gaussian distributions $\beta \sim N(0, 0.01)$, parameterized 259 by their precision (inverse of the variance). 260 In the third, and final, level of hierarchy, prior knowledge about the hyperparameters is 261 expressed. The hyperparameters are all derived from the spatial effect. More precisely, 262 we assume that W follows a prior Gaussian distribution with zero mean and covariance 263 matrix depending of hyperparameters κ and τ , which determine the range of the spatial 264 effect and the total variance, respectively. 265 As usual in this context, the resulting hierarchical Bayesian model has no closed 266 expression for the posterior distribution of all the parameters, and so numerical 267 approximations are needed. Here, we use the integrated nested Laplace approximation 268 (INLA) methodology [24] and software (http://www.r-inla.org) as an alternative to the 269 Markov chain Monte Carlo (MCMC) methods. The main reason for this choice is the 270 speed of calculation and the possibility of comparing many different models [24]. 271 Inference and prediction in unsampled locations were performed simultaneously by 272 INLA. For this, we used the Stochastic Partial Differential Equation module [29], which 273 allows us to fit the particular case of continuously indexed Gaussian Fields by INLA, as 274 is the case with our spatial component. Once the inference is carried out, the next step 275 is to predict the occurrence probability for paramphistomosis in the rest of the area of 276 interest (Galicia in our case), especially in non-observed locations. Following the

Bayesian reasoning, the occurrence of the species at new locations are considered as random variables, so that it is possible to obtain a set of probable values, together with the probabilities of them being the true values at each of those new specific locations. One way of computing the posterior predictive distributions for a whole region is to use the unobserved vertices of a triangulation of the region as prediction locations, as shown in Fig. 1. For the other points in the region of interest, the set of probable values are obtained by use of additional interpolation tools.

284 Models were compared by considering two criteria: the Deviance Information Criterion

[30], usually denoted as DIC, which is computed routinely by INLA as the default

286 criterion for comparing hierarchical models, and the Conditional Predictive Ordinate

287 (CPO), which has been used as a predictive measure (of the models). In particular, as

indicate by Roos and Held [31], we computed the mean logarithmic CPO (\overline{LCPO}).

Lower values for both DIC and *LCPO* indicate better models.

290 The posterior means and first and third quartiles from the predictive distribution were

291 plotted to illustrate the predicted probability of occurrence of infection.

292

293 Results

The necropsies revealed that 111 out of the 589 (18.8%; 95% Bayesian Credible

295 Interval (BCI): 15.7%-21.9%) cows harboured Paramphistomidae flukes in their rumens

296 or reticula. The parasite burdens per animal ranged between 1 and 11895 (median=

297 266) flukes, and the parasites were located in much higher proportion in the rumen

298 $(94.3\% \pm 14.1\%)$ than in the reticulum $(5.7\% \pm 8.3\%)$. The distribution of parasites

299 within the rumen was also heterogeneous, with higher percentages found in the atrium

300 (58.2% ± 28.6%) and the rumenoreticular sulcus (26.5% ± 20.0%) than in the ventral

and dorsal sacs (9.5% \pm 13.2% and 0.4% \pm 1.1%, respectively).

302 Breakdown of the results of necropsies and coprological analyses by production type

303 (beef, milk) and age (<7 years, between 7-9 years, > 9 years) revealed that the

304 prevalence of infection was higher in the beef cows than in dairy cows (29.2% 305 compared with 13.9%), although with widely variable fluke burdens in both types of 306 animals (Table 1). In both groups, stool analysis also revealed high percentages of 307 infected animals excreting widely varying amounts of eggs. With regard to age, in cows 308 aged over 9 years, the prevalence of infection (25.0%), parasite burdens (range=5-309 11895; median=351), percentage of animals with positive coprology, and faecal egg 310 counts (96.3 % cows excreting 1-2762 epg) were all higher than the younger cows. The 311 Bayesian GLM analysis (Table 2) showed that the effect of the type of production on 312 the prevalence of the infection, in particular the higher prevalence in beef cows, was 313 important as 95% BCI of the parameter for dairy cows clearly shifted towards negative 314 values. However, the type of production was not related to the fluke burdens, the 315 percentage of infected animals with positive coprology or the number of epg excreted 316 in faeces. The effect of age on the percentage of animals with positive coprology was 317 relevant, and the highest value was observed in cows over 9 years old (95% BCI of the 318 parameter shifted towards positive values). There was no relationship between the age 319 group and the prevalence of infection, the fluke burden or the number of epg. Finally, 320 there was a positive relation between the egg counts in the faeces and the parasite 321 burdens (posterior mean = 0.003; 95% CI = 0.002-0.004).

All the specimens examined were identified as *C. daubneyi*, on the basis of their
morphoanatomical characteristics. Moreover, the subsequent molecular analysis by
ITS-2 sequencing confirmed this specific classification, since the DNA amplification
revealed, in all specimens examined, a 410 bp fragment with a nucleotide composition
identical (100% homology) to that published for *C. daubneyi* in Gen BankTM
[GenBank: AY790883].

A total of 235 roe deer hunted in the vicinity of cattle farms distributed throughout
Galicia were examined by necropsy; no infections by paramphistomes were found in
any of them.

331 Not surprisingly, given the distribution of the cattle farms in Galicia (see Materials and 332 Methods 2.1), the dairy and beef farms sampled in this study were unevenly distributed 333 throughout the region (Fig. 2). Because of this and the large differences in the 334 prevalences of *C. daubneyi* infection in dairy and beef cows, we modelled the 335 probability of occurrence of this infection separately in the different types of cattle. The 336 model selected by the DIC and \overline{LCPO} criteria for fitting to the data on C. daubneyi 337 infection in dairy cows included the annual mean temperature and the log-transformed 338 slope (for smoothing the effect and preserve the linearity of this variable), as 339 covariates, and a stochastic spatial component that accounted for the residual spatial 340 autocorrelation (Table 3). The annual mean temperature had a negative effect on the 341 occurrence of C. daubneyi infection, whereas the effect of the slope on the response 342 variable was positive and more important (in the sense that the posterior probability of 343 being different from zero was greater). The spatial component showed a strong effect, 344 with positive values in the centre of Galicia (specifically in the southern part of the 345 provinces of A Coruña and Lugo), and values around zero in the northern, western and 346 southern zones of the study area (Fig. 3, which depicts the posterior mean (A) and 347 standard deviation (B) of the spatial random effect).

348 The maps included in Fig. 4 show the mean predicted distribution of the probability of 349 infection of dairy cows across Galicia (A) and the uncertainty around this estimation 350 provided by the lower 25% (B) and upper 75% (C) quartiles of the posterior distribution 351 for the predicted probability of infection by C. daubneyi. It should be noted that because 352 of the small population of dairy cattle in the south and east of Galicia, very few cows 353 from these areas were included in the sample (see points overlaid on Fig. 4A), so that 354 predictions obtained for those locations should be interpreted with caution. Excluding 355 these areas, it can be seen that the highest values of the probability of occurrence 356 of C. daubneyi infection covers the centre of Galicia, while the predicted probability for 357 the warmer and flatter western fringe is very low.

The results of inference for beef cows are shown in Table 4. In this case, the rainfall and the density of cows in the municipality of origin were incorporated in the model as covariates, and no spatial effect was detected. The probability of occurrence of infection in beef cows is higher in the central part of Galicia, where the municipalities with the highest densities of cattle are located (Fig. 5). The highest probabilities of infection (up to 0.8) were predicted for some small zones scattered within this central area, mainly in the farthest eastern zone, which is the driest zone within the region.

365

366 Discussion

367 This paper presents the results of a comprehensive study designed to investigate 368 different aspects of the epidemiology of bovine paramphistomosis in Galicia, the main 369 cattle-producing region in Spain. One interesting finding of the study is that infections 370 by paramphistomes are more prevalent among beef cows (29.2%) than among dairy 371 cows (13.9%), which probably reflects differences in exposure to infection, possibly 372 because of a closer association with pasture in the case of beef cows. This suggests 373 that the prevalence of parahimphistomosis has so far been underestimated in Galician 374 beef cattle. Earlier coprological surveys conducted in such animals had reported 375 prevalence rates of 10%-19% [32, 22, 23]. This discrepancy may be due to the different 376 origin of the animals examined. In the present study, prevalences were estimated from 377 data from animals on farms throughout Galicia, whereas only cattle from the northeast 378 of the region (province of Lugo) had previously been analyzed. Therefore, the higher 379 figure found in the present study may be a more accurate estimate of the true 380 prevalence of these infections in the whole region. Moreover, as demonstrated in the 381 present study, coprology does not detect all infections observed by necropsy. 382 Few studies have reported the size of the paramphistomes burdens present in naturally 383 infected cattle; however, the number of flukes in rumens and reticula determines the 384 degree of damage caused in the rumen and the potential detrimental effect on animal 385 health and production [4]. Most cows in the present study had low parasite burdens

(Median=266 flukes), similar to those previously reported by Arias et al. [33] for northwest Spain and northern Portugal and by Szmidt-Adjidé et al. [34] for central France.
Nevertheless, we also found some animals harbouring large parasite burdens (up to
11895 flukes), which might cause production losses, particularly under conditions of
stress [9].

391 Stool analysis revealed that a high proportion of infected cows (83.8%) excreted 392 detectable amounts of paramphistomid eggs in their faeces. Specifically, 96.3% of 393 cows over 9 years shed fluke eggs, sometimes in large amounts (up to 2762 epg). 394 These results highlight the importance of adult cows, especially older cows, in 395 maintaining and transmitting paramphistomosis within cattle herds. Adult cows, which 396 constitute the bulk of animals in herds, spend long periods of their life on pastures, 397 depositing large volumes of faeces that contaminate snail habitats with numerous 398 parasite eggs. Therefore, mature cows are important reservoirs that transmit the 399 infection to the new generations of the susceptible animals entering the herd. 400 Identification of wild reservoir hosts for parasites is a prerequisite for the control of 401 parasitic diseases in endemic areas. However, to date no study had investigated the 402 occurrence of natural infections by paramphistomes in roe deer, which is the most 403 abundant wild ruminant in Galicia and which may act as a reservoir for 404 paramphistomosis [35, 36]. In the present study, paramphistomid flukes were not found 405 in any of the 235 roe deer examined by necropsy, which clearly demonstrates that the 406 role of this animal species in the current epidemiology of paramphistomosis in this 407 region is negligible. The apparent absence of wild reservoir hosts for this parasitosis in 408 Galicia underlines the importance of adult cows as reservoir for the infection in this 409 region.

Paramphistomosis may be caused by different genera and species of the family
Paramphistomidae, which are difficult to identify due to the morphological similarities
among them. Indeed, most reports of paramphistomosis do not specify the species that
cause the disease. Nevertheless, species might differ in important aspects such as the

414 range of susceptible hosts, pathogenicity, resistance to drugs and the geographical 415 distribution, among other factors. Calicophoron daubneyi appears to be the most 416 common paramphistomid affecting cattle in Europe [15, 37]. This was also the only 417 species reported by Díaz et al. [32, 22, 23] and Arias et al. [33] in Galicia, although 418 neither the number of specimens identified nor the method used for identification were 419 specified in these studies. In the present study, we used morpho-anatomical (n=618) 420 and molecular techniques (n=82) to examine a large number of fluke specimens 421 collected from various anatomical zones within the rumens and reticula of animals from 422 different areas of Galicia. This methodology enabled us to confirm that C. daubneyi is 423 the only species of Paramphistomidae parasitizing cattle in Galicia. 424 As with other trematodes, the transmission of *C. daubneyi* depends on the distribution 425 of the intermediate host snails involved in its life cycle. This distribution, as well as the 426 development and survival of the intra-molluscan and free-living stages of the parasite 427 are determined by climatic and other environmental conditions, which may vary 428 considerably among the different locations within wider geographic areas. Therefore, 429 the spatial distribution of infection risk is not homogeneous, and estimates of its 430 geographical distribution are needed to identify the locations where the risk of infection 431 is high and where monitoring and control interventions should be targeted. Over the 432 last decade, several studies have used satellite-derived environmental data to model 433 the occurrence and prevalence of animal helminthosis [38, 39, 40, 41]. Although a 434 useful benchmark for future applications, these approaches could be improved by 435 applying more flexible and robust spatial statistical methods, such as those based on 436 Bayesian approach. These methods offer advantages over those based on traditional 437 statistics since they enable the spatial correlation of the variables and the uncertainty of 438 the parameters to be included in the modelling process. Although Bayesian analysis 439 has been increasingly incorporated to the study of the geographical distribution of 440 human helminthosis in recent years [42, 43, 44, 45, 46, 47, 48], it has scarcely been

441 used in the context of animal diseases [49, 50, 51], probably because of the relative 442 complexity and computational difficulty of Bayesian modelling [42]. 443 In the present study, spatial predictions of the probability of *C. daubneyi* infection 444 across Galicia were based on hierarchical Bayesian models developed separately for 445 dairy and beef cattle. The best models (in terms of DIC and \overline{LCPO}) suggested that 446 different covariates predicted the probability of infection in each type of cattle. The 447 spatial pattern in dairy cattle was mainly governed by temperature and slope in the 448 areas where the farms were located, so that the probability of infection increased with a 449 decreasing mean temperature and increasing slope. However, in beef cattle, the 450 annual rainfall and density of cows were the main predictive covariates, and infections 451 were more likely to occur in locations where the rainfall was lower and the stocking rate 452 was higher. The positive relationship between density of cows and probability of 453 infection was consistent with the general theoretical knowledge about pasture-454 transmitted parasitosis. However, different studies have demonstrated that the effects 455 of environmental covariates on the occurrence and/or prevalence of trematode 456 infections are not so obvious. In this regard, Raspch et al. [52] indicated that in the 457 case of Fasciola hepatica, a fluke that has the same snail as an intermediate host, the 458 risk of infection increases with increasing rainfall, until reaching a maximum at 459 approximately 90 mm per month; however, large amounts of rainfall (over 210 mm per 460 month) can wash parasite larvae and snails away and separate them from each other, 461 thus inhibiting transmission. Subsequently, Bennema et al. [53] suggested that the 462 threshold for such a "wash away" effect may be considerably lower, since they found a 463 negative association between rainfall and presence of economically important F. 464 hepatica infections in a region of Belgium (Flanders) where the average values for 465 annual and monthly rainfall were 800 mm and 61.1 mm, respectively. The present 466 results in a zone with a similar mean annual precipitation (1065 mm) appear to confirm 467 this suggestion. As regards temperature, it is known that, under laboratory conditions 468 (i.e., with sufficient moisture) the development rate of the fluke larval stages increases

469 between 10 °C and 25 °C [54], so that temperatures within this range are also 470 expected to be a risk factor for infection of the ruminants. However, under field 471 conditions, the relationship between risk of infection and temperature is more complex, 472 largely due to the strong interactions between air temperature, moisture content of 473 ground and soil permeability. The negative association between temperature and 474 probability of infection in the present study suggests that the suitable thermal range for 475 transmission of infection in Galicia is narrower than expected, probably because most 476 soils in this region are highly permeable and dry out quickly in summer, when the 477 temperature increases in the absence of rainfall. Therefore, in the warmest locations, 478 interruption of the emission of cercariae and shortening of survival of metacercariae will 479 occur during summer, with the subsequent reduction in the intensity of infection 480 transmission. In the present study, slope was positively associated with the probability 481 of infection. This is consistent with the findings reported by Cringoli et al. [55] for C. 482 daubneyi and by Bennema et al. [53] for F. hepatica infections, which the authors 483 attributed to the ponding (temporary or permanent) that typically occurs in the lower 484 areas of sloping pastures in rainy locations. In contrast, McCann et al. [56] found a 485 negative effect of the slope on the prevalence of F. hepatica infection, which they 486 attributed to better drainage. In a study carried in north-eastern Galicia, Díaz et al. [22] 487 also found a negative effect of slope on the prevalence of *C. daubneyi* infection, 488 although only when the slope was very steep (exceeding 25%), so that the previously 489 observed discrepancies may be due to differences in the ranges of slope values in the 490 different studies. However, the results of most previous studies should be regarded 491 with caution due to the unsuitability of the applied statistical tests (chi-square test, 492 Spearman rank order correlation and linear regression), which may have led to biased 493 parameter estimates as spatial correlation was ignored. 494 There were two important differences in the models fitted for the probability of C. 495 daubneyi infection in dairy and beef cattle. First, different environmental covariates

496 were used to explain the probability of the infection in both models. This may reflect

497 variations in the effects of environmental covariates depending on the range of values 498 and potential interactions among them in the different areas where the farms of origin 499 of both types of animals were located. In this sense, it must be taken into account the 500 dairy and beef farms were unevenly distributed throughout the study area (see 501 Materials and Methods) and that values for covariates at the locations of dairy and beef 502 farms differed significantly (data not shown). The second difference is related to the 503 residual spatial correlation, which existed in the model of infection transmission of dairy 504 cows, but not in that of beef cows. The overall transmission success depends on the 505 establishment of the parasite in the ruminant host, which in turn is determined by the 506 ingestion of infective metacercariae during grazing. Since beef cows in Galicia graze 507 throughout the year and throughout their lives, infection is guaranteed when the grass 508 is infected with metacercariae. Therefore, variations in the risk of infection of beef cattle 509 will be primarily determined by the environmental covariates that determine 510 development of the parasite, its intermediate host, and ultimately the presence of 511 infective metacercariae on the grass. However, the grazing patterns of dairy cows are 512 different: they may graze for their whole life or only during some stages (heifers, dry 513 cows), throughout the whole year or only seasonally, or they may even never graze on 514 pasture. This implies great differences as regards the possibilities of contact with 515 metacercariae and in the actual risk of infection. As we could not include this 516 information in the modelling process, the residual autocorrelation of the infection in 517 dairy cattle may reflect the risk of infection associated with the differences in the 518 grazing history of the animals. 519 This study has provided the first maps of the spatial distribution of the probability of C.

daubneyi infection in beef and dairy cattle. Its use allowed us to identify the centre of Galicia as the zone with the highest risk of infection for both types of cattle. This is also the zone with the largest livestock population, so that it should be the preferential target for campaigns to raise the awareness of farmers and veterinarians about the situation and for monitoring interventions. Nevertheless, in the case of dairy cows, the estimates

525 may be inaccurate because of the lack of information about grazing patterns and other 526 management factors which were not measured in this study. For example, de-worming 527 treatments will affect the establishment, survival and fecundity of adult worms, and 528 therefore will also affect the transmission patterns of the infection. Consequently, the 529 next logical step is to include such information to further refine the model and enhance 530 the accuracy of prediction. Furthermore, we believe that the use of this approach for 531 constructing maps of the spatial distribution of infections and co-infections by other 532 common helminths in the region, such as those produced by *F. hepatica* and 533 gastrointestinal nematodes, may help design integrated programs for more efficient 534 surveillance and control of bovine helminthoses.

535

536 Competing interests

- 537 The authors declare that they have no competing interests.
- 538

539 Authors' contributions

540 Conception and design of the study: MGW and MM. Sample and data collection: MGW,

541 JACH and MM. Species identification by microscopy and molecular techniques: AMMI

and YMG. Bayesian statistical analysis: SL, DC, FM, ALQ. Manuscript draft

543 preparation: MGW, JACH, DC, YMG and MM. All authors have read and approved the

544 final manuscript.

545

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Figure 1. Triangulation map covering the region. Each mesh vertex is either an
observed point (•) or a prediction point.

792

Figure 2. Geografical distribution of the farms of origin of the dairy cows (A) and
beef cows (B) sampled in the slaughterhouse. Cows infected by *C. daubneyi* (•)
and uninfected cows (•).

796

Figure 3. Spatial component of the fitted model for the probability of occurrence
of *Calicophoron daubneyi* infection in dairy cows throughout Galicia. Posterior
mean (A) and standard deviation (B).

800

Figure 4. Predicted probability of the occurrence of *C. daubneyi* infection in

dairy cows throughout Galicia. Mean (A), $Q_{0.25}$ (B) and $Q_{0.75}$ (C) of the posterior distribution. The overlaid points mark the geographical origin of the cows in the sample, distinguishing between cows infected by *C. daubney*i (•) and uninfected cows (•).

806

Figure 5. Predicted probability of the occurrence of *C. daubneyi* infection in beef cows throughout Galicia. Mean (A), $Q_{0.25}$ (B) and $Q_{0.75}$ (C) of the posterior distribution. The overlaid points mark the geographical origin of the cows in the sample, distinguishing between cows infected by *C. daubneyi* (•) and uninfected cows (•).

812

Table 1 Prevalence and intensity of infections by paramphistomes in cows.

8	1	4
0		

		Ne	ecropsy	Coprology		
_	Ν	Prevalence (%)	Fluke burden Range (median)	Positive ¹ (%)	epg Range (median)	
Type of production						
Beef	192	29.2	3-11070 (257)	89.3	1-855 (16)	
Milk	397	13.9	1-11895 (307)	78.2	1-2762 (31)	
Age (years)						
<7	200	14.5	4-6009 (305)	79.3	2-1100 (18)	
7-9	173	16.2	1-4664 (165)	64.3	3-573 (19)	
>9	216	25.0	5-11895 (351)	96.3	1-2762 (25)	
Total	589	18.8	1-11895 (266)	83.8	1-2762 (22)	

¹ Percentage calculated on the basis of the number of animals with infection by paramphistomes detected by necropsy

Table 2 Bayesian GLM and LM analysis of results obtained by necropsy and coprological analysis.

	Posterior distribution means (95% BCI) ¹			
-	Prevalence	Fluke burden	Positive coprology	epg
Intercept including reference				
group Beef and <7 years Type of production	-1.13 (-1.67, -0.61)	5.33 (4.35, 6.30)	1.57 (0.41, 2.89)	0.88 (0.84, 0.92)
Milk Age (vears)	-0.83 (-1.30, -0.36)	0.26 (-0.59, 1.11)	-0.29 (-1.48, 0.85)	-0.01 (-0.05, 0.02)
7-9	0.12 (-0.45, 0.69)	-0.85 (-1.96, 0.26)	-0.80 (-2.02, 0.36)	0.03 (-0.02, 0.07)
>9	0.33 (-0.21, 0.88)	0.35 (-0.67, 1.36)	1.84 (0.25, 3.70)	-0.03 (-0.08, 0.01)
DIC ²	557.76	489.24	90.67	-220.43

820 BCI: Bayesian credible interval;² DIC: Deviance information criterion

Table 3 Bayesian spatial logistic regression model for the probability of infection of dairy cows with *C. daubneyi*. 822

	Summary of the posterior distributions ¹						
		Mean	SD	Q _{0.025}	Q _{0.5}	Q _{0.975}	
	(Intercept)	1.259	2.587 -	-3.852	1.270	6.308	
	Temperature	-0.398	0.233 -	-0.853	-0.398	0.061	
	Slope (Log)	0.517	0.228	0.082	0.512	0.979	
823	DIC=305.04; LCP	O=0.3849					
824	¹ Coefficients were	calculated t	aking into a	ccount the	e geospatial	effect.	
825	25						
826							
827							
828 Table 4 Bayesian spatial logistic regression model for the probability of infection of beef cows with				ility of infection of beef cows with C. daubneyi.			
829		·	•			•	
	Summary of the posterior distributions						
		Mean	SD	Q _{0.025}	Q _{0.5}	Q _{0.975}	-
	(Intercept)	0.510	1.183	-1.791	0.503	2.850	
	Rainfall	-0.003	0.001	-0.005	-0.003	-0.000	
	Cattle density	0.040	0.009	0.022	0.039	0.058	

830 831 DIC=209.96; LCPO=0.55





B)





B)





A)

B)





C)



A)

B)





